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<p>(54) Title: METHODS FOR THE DIAGNOSIS AND TREATMENT OF METASTATIC PROSTATE TUMORS (57) Abstract <p>The present invention is directed to methods for the identification of a prostate cancer cell that has metastatic potential or a cell that is or is derived from a secondary prostate tumor metastasis by screening for the expression of flt-4, the cellular receptor of vascular end thelial growth factor-C and -D ("VEGF-C", "VEGF-D"). The present invention is also directed to methods for treating, inhibiting or preventing secondary prostate tumor metastases by inhibiting the expression or activity of flt-4, e.g., inhibiting flt-4: VEGF-C/D complex formation (binding), by administration of a therapeutic. Compositions useful in such methods are also provided.</p></p>		

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METHODS FOR THE DIAGNOSIS AND TREATMENT OF METASTATIC PROSTATE TUMORS

1. FIELD OF THE INVENTION

The present invention is directed to methods for the identification of a prostate cancer cell that has metastatic potential or a cell that is a secondary prostate tumor metastasis, or is derived therefrom, by screening for the expression or activity of flt-4, the cellular receptor of vascular endothelial growth factor-C ("VEGF-C") and -D ("VEGF-D").

- 10 The present invention is also directed to methods for treating, inhibiting or preventing secondary prostate tumor metastases by inhibiting the expression or activity of flt-4, e.g., inhibiting flt-4:VEGF-C complex or flt-4-VEGF-D complex formation (binding). Compositions useful in such methods are also provided.

2. BACKGROUND OF THE INVENTION

The development and progression of prostate cancer remains poorly understood. Neoplastic transformation involves multiple mechanisms, such as p53 or ras mutations, and/or imbalance in growth regulatory factors.

- VEGF is a heparin-binding, dimeric polypeptide growth factor originally
20 purified based on its vascular permeability- enhancing activity and subsequently shown to be a potent mitogen for endothelial cells (reviewed in Shibuya, 1995, Adv. Cancer Res. 67:281-316). VEGF influences both angiogenesis and vascular permeability in solid tumors (Ferrara, 1995, Breast Cancer Res. Treat. 36:127-137; Ferrara et al., 1991, J. Cell Biochem. 47:211-8.; Klagsbrun et al., 1996, Cytokine Growth Factor Rev. 7:259-70). It is also
25 produced by some normal tissues, particularly during the menstrual cycle (Shweiki et al., 1993, J. Clin. Invest. 91:2235-2243) and wound healing (Frank et al., 1995, J. Biol. Chem. 270:12607-12613), and is thought to play important role(s) in the angiogenic pathology of diabetic retinopathy and rheumatoid disorders (Ferrara, 1995, Breast Cancer Res. Treat. 36:127-137).

- 30 In the prostate, the expression of VEGF has been reported both in non-malignant epithelium (Brown et al., 1995, J. Urol. 154:576-579; Jackson et al., 1997, J. Urol. 157:2323-2328) as well as in cancer cells (Ferrer et al., 1997, J. Urol. 157:2329-2333; Ferrer et al., 1998, Urology 51:161-167; Harper et al., 1996, Br. J. Cancer 74:910-916; Jackson et al., 1997, J. Urol. 157:2323-2328). Localization studies showed the presence of
35 VEGF, at both mRNA and protein levels, in prostate cancer cells, tumor-associated stroma, and a proportion of benign prostate hyperplasia ("BPH") epithelial cells (Jackson et al., 1997, J. Urol. 157:2323-2328). The relative level of VEGF in normal and transformed

prostate epithelium remains controversial and unclear (Woessner et al., 1998, *Exp. Mol. Pathol.* 65:37-52; Chevalier, 1997, *J. Urol.* 157:2040-2041). In prostate cancer, increased microvessel density was found to correlate with disease stage and metastasis (Bigler et al., 1993, *Hum. Pathol.* 24:220-6; Brawer et al., 1994, *Cancer* 73:678-687.; Deering et al., 1995, *Prostate* 26:111-115.; Siegal et al., 1995, *Cancer* 75:2545-2451; Weidner et al., 1996, *Important Adv. Oncol.* 167-190.). Increased neo-vascularization is also found in prostatic intraepithelial neoplasia ("PIN") (Brawer et al., 1994, *Cancer* 73:678-687; Ferrer et al., 1997, *J. Urol.* 157:2329-2333; Ferrer et al., 1998, *Urology* 51:161-167; Harper et al., 1996, *Br. J. Cancer* 74:910-916) and latent carcinoma (Furusato et al., 1994, *Br. J. Cancer* 70:1244-1246). Elevated levels of angiogenesis are found in human cell lines PC-3 and DU-145-derived tumors when implanted in nude mice (Connolly et al., 1998, *J. Urol.* 160:932-936). Taken together, these studies suggest a trophic role for VEGF in supporting tumor growth via angiogenesis.

VEGF-C is another endothelial growth factor with 32% amino acid identity to VEGF and was originally cloned from a human prostate hormone refractory cell line, PC-3 (Joukov et al., 1997, *J. Cell Physiol.* 173:211-215; Joukov et al., 1996, *EMBO J.* 15:290-298; Joukov et al., 1997, *EMBO J.* 16:3898-3911). Whereas VEGF is specific to endothelial cells of blood vessels, VEGF-C is a lymphangiogenic factor. The receptors for VEGF-C, both flk-1 and flt-4, are expressed in lymphatic endothelial cells and melanocytes (Kaipainen et al., 1993, *J. Exp. Med.* 178:2077-2088). Experiments with transgenic mice showed that the expression of VEGF-C is associated with the development of lymphatic vessels in normal tissues (Jeltsch et al., 1997, *Science* 276:1423-1425). VEGF-C induced growth of lymphatic vessels in chick chorioallantoic membrane (Oh et al., 1997, *Dev. Biol.* 188:96-109).

Flt-4 is a receptor type tyrosine kinase (RTK) with 7 Ig-like domains similar to other VEGF receptors. Expression of flt-4 was initially localized to angioblasts and venules in the early embryo. It becomes restricted to lymphatic endothelial cells during later development, and some high endothelial venules in human adult tissues (Kaipainen et al., 1995, *Proc. Natl. Acad. Sci. USA* 92:3566-3570; Kaipainen et al., 1993, *J. Exp. Med.* 178: 2077-2088). Expression of Flt-4 is consistent with known lymphatic vascular pattern in human skin (Lymboussaki et al., 1998, *Am. J. Pathol.* 153: 395-403). This expression pattern suggested that Flt-4 may play a role in the regulation of lymphangiogenesis (Kaipainen et al., 1995, *Proc. Natl. Acad. Sci. USA* 92:3566-3570; Kaipainen et al., 1993, *J. Exp. Med.* 178: 2077-2088; Mustonen et al., 1995, *J. Cell Biol.* 129: 895-898). Expression of Flt-4 has also been found in melanocytes (Gitay-Goren et al., 1993, *Biochem. Biophys. Res. Commun.* 190:702-708). In addition to VEGF-C, VEGF-D is also a ligand for flt-4.

(Achen et al., 1998, Proc. Natl. Acad. Sci. USA 95:548-553; Yamada et al., 1997, Genomics 42:483-488).

Citation or identification of any reference in Section 2 or any other section of this application shall not be construed as an admission that such reference is available as prior art to the present invention.

3. SUMMARY OF THE INVENTION

The present invention is based, in part, on the surprising discovery that the expression of flt-4 in a prostate cell indicates that such prostate cell is a cancerous prostate cell that has metastatic potential or is a secondary tumor metastasis of a primary prostate tumor, or is derived therefrom.

The present invention is directed to methods for detection of metastatic potential comprising detecting expression or activity of flt-4 in a prostate cancer cell, wherein expression or activity of flt-4 indicates that said cell has metastatic potential. The present invention is also directed to methods for detection of metastatic potential comprising identifying a prostate cell in a body fluid sample obtained from a subject and detecting expression or activity of flt-4 in said cells, wherein expression of flt-4 indicates that said cell is a prostate cancer cell that has metastatic potential or is a secondary tumor metastasis or is derived therefrom. The prostate cell can be identified using a prostate cell-specific marker and can be obtained from a body fluid sample, e.g., blood, urine, semen. In a preferred embodiment, the prostate cell can be identified and screened for flt-4 expression simultaneously using an antibody specific for a prostate cell-specific marker and an antibody specific for flt-4 in a flow cytometer. In another preferred embodiment, the prostate cell can be identified and screened for flt-4 expression simultaneously using an antibody specific for a prostate cell-specific marker and an antibody specific for flt-4 using a laser scanning cytometer.

The present invention is also directed to methods for diagnosing metastatic prostate cancer in a subject comprising identifying a prostate cell in a body fluid sample obtained from the subject and detecting expression or activity of flt-4 in the prostate cell, wherein expression or activity of flt-4 in said cell indicates that the subject has metastatic prostate cancer. The present invention is also directed to methods for determining the prognosis of a subject with prostate cancer comprising identifying a prostate cell in a body fluid sample obtained from a subject with prostate cancer and detecting expression or activity of flt-4 in the prostate cell, wherein expression or activity of flt-4 in the prostate cell obtained from the subject indicates that the subject has a worse prognosis as compared to a second subject in whose prostate cell no flt-4 expression or activity is detected. Methods

for monitoring the efficacy of a method of treatment or of inhibition of metastatic prostate cancer are also provided.

The present invention is directed to methods for treating, inhibiting or preventing a secondary prostate tumor metastasis in a subject by administering one or more compounds or molecules that inhibits flt-4 expression or activity. Illustrative examples of such compounds or molecules include, but are not limited to, fragments of flt-4 or nucleic acid molecules encoding the same, antisense oligonucleotides to inhibit expression of flt-4, antibodies specific to flt-4 or to a complex of flt-4 and its ligand VEGF-C or its ligand VEGF-D, etc. The present invention is further directed to compositions which can be used in the above described methods.

The present invention is further directed to methods for inhibiting the activity of a flt-4:VEGF-C complex or flt-4:VEGF-D complex. Methods that identify molecules that inhibit flt-4 expression or activity are also provided. Animal models and methods for screening for modulators (*i.e.*, agonists and antagonists) of the expression or activity of flt-4 or the activity of a flt-4:VEGF-C or flt-4:VEGF-D complex are also provided.

4. BRIEF DESCRIPTION OF THE FIGURES

Figs. 1A-1F set forth the nucleotide sequence (SEQ ID NO:1) and the amino acid sequence (SEQ ID NO:2), respectively, of the human flt-4 gene.

Figs. 2A-2F are photographs of anti-flt-4 antibody-stained prostatic tissue sections. Fig. 2A: normal lymph node; Fig. 2B: benign prostatic hyperplasia tissue; Fig. 2C, 2D: lymph node with prostatic metastases; Fig. 2E: benign prostatic hyperplasia tissue and prostate cancer; and Fig. 2F: prostate cancer.

Figs. 3A-3D are photographs of anti-flt-4 antibody stained human prostate cell lines. Fig. 3A: LNCaP cell line; Fig. 3B: PC3 cell line; Fig. 3C: DU145; and Fig. 3D: TSUPr1 cell line.

Fig. 4 is a photograph of agarose gels showing the results of reverse transcriptase polymerase chain reactions ("RT-PCR") testing for expression of VEGF, VEGF-C and their receptors flk-1 and flt-4, respectively, in several human prostate cancer cell lines.

Fig. 5 is a graph showing that flt-4 expression can be detected on the surface of live LNCaP cells using an anti-flt-4 antibody.

Fig. 6 is a histogram showing the results of antisense oligonucleotide treatment of a human prostate cancer cell line.

5. DETAILED DESCRIPTION OF THE INVENTION

The present invention is based, in part, on the surprising discovery that the expression of flt-4 in a prostate cell indicates that such prostate cell is a cancerous prostate cell that has metastatic potential or is a secondary tumor metastasis of a primary prostate tumor, or is derived therefrom. Not intending to be limited to a particular mechanism of action, although it was known that VEGF-C and its cell surface receptor flt-4 are implicated in the promotion of lymphoangiogenesis, prior to the present invention, it was not known that VEGF-C and flt-4 are involved in an autocrine feedback loop resulting in the promotion and formation of secondary prostate tumor metastases. Further, the present inventor believes that VEGF-D, another ligand of flt-4, and flt-4 are also involved in an autocrine feedback loop resulting in the promotion and formation of secondary prostate tumor metastases.

The present invention is directed to methods for detection of metastatic potential comprising detecting expression or activity of flt-4 in a prostate cancer cell, wherein expression or activity of flt-4 indicates that said cell has metastatic potential. The present invention is also directed to methods for detection of metastatic potential comprising identifying a prostate cell in a body fluid sample obtained from a subject and detecting expression or activity of flt-4 in said cells, wherein expression of flt-4 indicates that said cell is a prostate cancer cell that has metastatic potential or is a secondary tumor metastasis, or is derived therefrom. The prostate cell can be identified using a prostate cell-specific marker and can be obtained from a body fluid sample, *e.g.*, blood, urine, semen. In a preferred embodiment, the prostate cell can be identified and screened for flt-4 expression simultaneously using an antibody specific for a prostate cell-specific marker and an antibody specific for flt-4 in a flow cytometer.

Methods for treating, inhibiting or preventing secondary prostate tumor metastases by administration to a subject in need of such treatment, inhibition or prevention by administration of a molecule that inhibits the expression or activity of flt-4 are provided. Examples of such molecules include, but are not limited to, a fragment of flt-4 that acts as a competitive inhibitor of flt-4 binding to its ligands VEGF-C or VEGF-D, antibodies to flt-4, antisense nucleic acids, etc.. Methods for the diagnosis, prognosis and screening for secondary prostate tumor metastases are also provided. Compositions for carrying out such methods are also provided in the present invention. For example, such methods can be used by a pathologist screening tissue biopsies, such as prostate tissue or lymph node tissue biopsies, for a prostate cell with metastatic potential.

The present invention is also directed to methods of assaying for the presence of a flt-4:VEGF-C complex or a flt-4:VEGF-D complex ("flt-4:VEGF-C/D") for

diagnosis and/or prognosis of prostate cancer. As used in the present application "flt-4:VEGF-C/D" indicates a complex of flt-4 and VEGF-C or a complex of flt-4 and VEGF-D.

For clarity of disclosure, and not by way of limitation, a detailed description of the invention is divided into the following subsections.

5.1. FLT-4 EXPRESSION IN PROSTATE CELLS

Flt-4 expression in a prostate cancer cell indicates that the prostate cancer cell has metastatic potential or indicates that the prostate cancer cell is a secondary prostate tumor metastasis, or is derived therefrom, and thus flt-4 expression has diagnostic and prognostic utility in assessing the metastatic potential of a prostate cancer cell or in screening for the presence of secondary tumor metastases derived from a primary prostate tumor.

In one embodiment of the invention, prostate cells are identified by expression of a prostate cell-specific marker and the identified prostate cells are screened for flt-4 expression. In another embodiment of the invention, the prostate cells are identified and screened concurrently with labeled antibodies specific for a prostate cell-specific marker and specific for flt-4, respectively, using a flow cytometer.

Detecting levels of flt-4:VEGF-C/D complexes, or flt-4 protein expression, or detecting the level of mRNAs encoding flt-4 in a prostate cell, may be used to determine whether the prostate cell has metastatic potential or whether the prostate cell is a secondary prostate tumor metastasis, or is derived therefrom. Moreover, the detection of flt-4:VEGF-C/D complexes or flt-4 expression in a prostate cell can be used in the prognosis of prostate cancer, to follow the course of prostate cancer, to follow a therapeutic response, etc.

Prostate cells can be detected by any method known to those of skill in the art. For example, prostate cells can be detected using an antibody specific for a prostate cell-specific marker or using a fragment comprising the binding domain of an antibody specific for a prostate cell marker. Preferably, the antibodies used are monoclonal antibodies.

Prostate cell-specific markers are known and include prostate-specific antigen (PSA), prostate-specific membrane antigen (PSMA), prostate secretory protein (PSP), prostate acid phosphatase (PAP), and human glandular kallekrein 2 (HK-2). Another prostate cell-specific marker is prostate stem cell antigen (PSCA) identified by Reiter et al., 1998, Proc. Natl. Acad. Sci. USA 95:1735-1740. Yet another prostate cell-specific marker is PTI-1, a prostate carcinoma oncogene identified by Shen et al., 1995, Proc. Natl. Acad.

Sci. USA 92:6778-6782. For a general review of prostate cell-specific markers, see Nelson et al., 1998, *Genomics* 47:12-25.

In a preferred embodiment, the prostate cells are detected by using a monoclonal antibody specific for PSMA. PSMA is an approximately 120 kDa molecular weight protein expressed in prostate tissues and was originally identified by reactivity with a monoclonal antibody designated 7E11-C5. Horoszewicz et al., 1987, *Anticancer Res.* 7:927-935; U.S. Patent No. 5,162,504. PSMA was obtained in purified form (Wright et al., 1990, *Antibody Immunoconjugates and Radio Pharmaceuticals* 3:Abstract 193) and characterized as a type II transmembrane protein having some sequence identity with the transferrin receptor (Israeli et al., 1994, *Cancer Res.* 54:1807-1811) and with NAALADase activity (Carter et al., 1996, *Proc. Natl. Acad. Sci. USA* 93:749-753). More importantly, although PSMA is expressed in normal prostate, benign prostate hyperplasia and prostate cancer, PSMA is expressed in increased amounts in prostate cancer, and an elevated level of PSMA is also detectable in the sera of these prostate cancer patients. Horoszewicz et al., 1987, *supra*; Rochon et al., 1994, *Prostate* 25:219-223; Murphy et al., 1995, *Prostate* 26:164-168; and Murphy et al., 1995, *Anticancer Res.* 15:1473-1479. A cDNA encoding PSMA has been cloned. Israeli et al., 1993, *Cancer Res.* 53:227-230.

PSMA-specific antibodies are known and include 7E11.C5 (ATCC Accession No. HB10494); 3F5.4G6 (ATCC Accession No. HB12060); 3D7-1.1 (ATCC Accession No. HB12309); 4E10-1.14 (ATCC Accession No. HB12310); 1G3 (ATCC Accession No. HB-12489); 1G9 (ATCC Accession No. HB-12495); 2C7 (ATCC Accession No. HB-12490); 3C4 (ATCC Accession No. HB-12494); 3C6 (ATCC Accession No. HB-12491); 3C9 (ATCC Accession No. HB-12484); 3E6 (ATCC Accession No. HB-12486); 3E11 (ATCC Accession No. HB-12488); 3G6 (ATCC Accession No. 12485); 4D4 (ATCC Accession No. HB-12493); 4D8 (ATCC Accession No. HB-12487); or 4C8B9 (ATCC Accession No. HB-12492), see WO 97/35616. Other PSMA-specific antibodies which can be employed in the present invention include E99 (ATCC Accession No. HB-12101); J415 (ATCC Accession No. HB-12109); J533 (ATCC Accession No. HB-12127); and J591 (ATCC Accession No. HB-12126), each isolated by Bander, International Patent Publication WO 98/03873. Additional non-limiting examples of antibodies specific for PSMA, which can be used in the methods of the present invention, are presented in Table I. All the antibodies presented in Table I are murine IgG monoclonal antibodies, which are reactive to native PSMA. The approximate location of the binding epitope of each antibody, as well as the isotype subclass of each antibody, is also summarized in Table I.

TABLE I
 Binding Specificity and Isotype of PSMA-Specific
 Antibodies to Native PSMA and PSMA Fragments

Antibody	Native PSMA	1-173	134-437	437-750	Isotype ^a
3F6	+	-	-	-	IgG _{2b}
2E4	+	weak	-	-	IgG _{2a}
3C2	+	+	+	-	IgG _{2a}
4C8G8	+	-	+	-	IgG _{2b}
2C4	+	-	+	-	IgG ₁
4C11	+	-	+	-	IgG ₁
1D11	+	-	+	-	IgG _{2b}
4E8	+	-	+	-	IgG _{2b}
2G5	+	-	+	-	IgG _{2b}
4E6	+	-	+	-	IgG ₁
1F4	+	-	+	-	IgG ₁
2E3	+	-	-	+	IgG _{2a}
3D8	+	-	-	+	IgG _{2a}
4F8	+	-	-	+	IgG _{2a}
3D2	+	-	-	+	IgG _{2a}
1G7	+	-	-	+	IgG _{2a}
3D4	+	-	-	+	IgG _{2a}
4D4	+	-	-	-	IgG ₁
5G10	+	-	+	-	IgG ₁
5E9	+	-	+	-	IgG ₁

^a Isotype specificity was determined using IsoStrip tests (Boehringer-Mannheim) for murine antibody isotype determinations which were conducted according to manufacturer's instructions.

In another embodiment, prostate cells are detected using a monoclonal antibody specific for a prostate cell-specific marker selected from the group consisting of prostate-specific antigen (PSA), prostate secretory protein (PSP), prostate acid phosphatase (PAP), human glandular kallekrein 2 (HK-2), prostate stem cell antigen (PSCA), and PTI-1.

In yet another embodiment, prostate cells are detected using the PR-1 monoclonal antibody (ATCC Accession No. HB-11145; Pastan, U.S. Patent No. 5,489,525).

Further, a portion of an antibody specific for a prostate cell marker, including purified fragments of the monoclonal antibodies having at least a portion of an antigen binding region, including such as Fv, F(ab')₂, Fab fragments (Harlow and Lane, 1988,

Antibody, Cold Spring Harbor), single chain antibodies (U.S. Patent 4,946,778), chimeric or humanized antibodies (Morrison *et al.*, 1984, Proc. Natl. Acad. Sci. USA 81:6851; Newuberger *et al.*, 1984 Nature 81:6851) and complementarity determining regions (CDR) can be used in the present invention. Mimetics of the antibodies can also be used in the present invention.

The prostate cell marker-specific antibody or portion thereof can, in turn, be detected by having the antibody labeled directly or indirectly with a detectable marker. Alternatively, the prostate cell marker-specific antibody ("first antibody") can be contacted with a second antibody which is specific for the prostate cell marker-specific antibody.

- 10 This second antibody can be labeled directly or indirectly with a detectable marker. Such detectable markers include, but are not limited to, a radioactive moiety, a substrate converting enzyme, fluorescent marker, biotin, and the like. For a general review, see, Antibodies, A Laboratory Manual, 1988, Harlow et al., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.

- 15 In an alternative embodiment, prostate cells can be detected by measuring the levels of nucleic acids and related nucleic acid sequences and subsequences, including complementary sequences, encoding a prostate cell-specific marker. For example, a cDNA encoding PSMA has been cloned. Israeli et al., 1993, Cancer Res. 53:227-230. The prostate cell-specific marker nucleic acid sequences, or subsequences thereof, comprising at least 8 nucleotides, can be used as hybridization probes. Hybridization assays can be used to detect the expression of the prostate cell-specific marker, and thus, detect prostate cells. In particular, such a hybridization assay is carried out by a method comprising contacting a sample containing nucleic acid with a nucleic acid probe capable of hybridizing to PSMA, under conditions such that hybridization can occur, and detecting or measuring any resulting hybridization.

- 25 Once the prostate cell has been identified, the prostate cell is screened for flt-4 expression. Flt-4 expression can be detected by any method known to those of skill in the art. For example, flt-4 expression can be detected using an antibody specific for flt-4 or using a fragment comprising the binding domain of an antibody specific for flt-4. Such antibodies are well known in the art and can be produced using methods well known in the art without undue experimentation, see Section 5.2.3, *infra*. In a preferred embodiment, flt-4 expression is detected using the anti-flt 4 antibody obtained from Santa Cruz Corp. (Santa Cruz, CA). Further, a portion of an antibody specific for a prostate cell marker, including purified fragments of the monoclonal antibodies having at least a portion of an antigen binding region, including such as Fv, F(ab')₂, Fab fragments (Harlow and Lane, 1988, Antibody, Cold Spring Harbor), single chain antibodies (U.S. Patent 4,946,778), chimeric

or humanized antibodies (Morrison *et al.*, 1984, Proc. Natl. Acad. Sci. USA 81:6851; Newuberger *et al.*, 1984 Nature 81:6851) and complementarity determining regions (CDR) can be used in the present invention. Mimetics of the antibodies can also be used in the present invention.

5 The immunoassays which can be used include but are not limited to competitive and non-competitive assay systems using techniques such as Western blots, radioimmunoassays, ELISA (enzyme linked immunosorbent assay), "sandwich" immunoassays, immunoprecipitation assays, precipitin reactions, gel diffusion precipitin reactions, immunodiffusion assays, agglutination assays, complement-fixation assays,
10 immunoradiometric assays, fluorescent immunoassays, protein A immunoassays, to name but a few known in the art.

 The flt-4 antibody or portion thereof can, in turn, be detected by having the antibody labeled directly or indirectly with a detectable marker. Alternatively, the flt-4 antibody ("first antibody") can be contacted with a second antibody which is specific for the
15 flt-4 antibody. This second antibody can be labeled directly or indirectly with a detectable marker. Such detectable markers include, but are not limited to, a radioactive moiety, a substrate converting enzyme, fluorescent marker, biotin, and the like. For a general review, see, Antibodies, A Laboratory Manual, 1988, Harlow et al., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.

20 Flt-4 expression can also be detected by measuring the levels of nucleic acids and related nucleic acid sequences and subsequences, including complementary sequences, encoding flt-4. A cDNA encoding flt-4 has been cloned (Galland et al., 1992, Genomics 13(2):475-478) and the nucleotide and amino acid sequences of flt-4 are set forth in Figs. 1A-F (SEQ ID NOS:1 and 2). The flt-4 nucleic acid sequences, or subsequences thereof,
25 comprising at least 8 nucleotides, can be used as hybridization probes. Hybridization assays can be used to detect the expression of flt-4 in a prostate cell, and thus, detect prostate cells with metastatic potential. In particular, such a hybridization assay is carried out by a method comprising contacting a sample containing nucleic acid with a nucleic acid probe capable of hybridizing to flt-4, under conditions such that hybridization can occur, and
30 detecting or measuring any resulting hybridization.

 In a preferred embodiment of the invention, the flt-4 expressing prostate cells can be detected using a flow cytometer. Fluorescence activated cell sorting (FACS) flow cytometry is a common technique for antibody based cell detection and separation. Typically, detection and separation by flow cytometry is performed as follows. A sample
35 containing the cells of interest is contacted with fluorochrome-conjugated antibodies, which allows for the binding of the antibodies to the specific-cell marker, such as PSMA and flt-4.

The bound cells are then washed by one or more centrifugation and resuspension steps. The cells are then run through a FACS which separates the cells based on the different fluorescence characteristics imparted by the cell-bound fluorochrome. FACS systems are available in varying levels of performance and ability, including multicolor analysis which is preferred in the present invention. For a general review of flow cytometry, see Parks et al., 1986, Chapter 29: Flow Cytometry and fluorescence activated cell sorting (FACS) in: Handbook of Experimental Immunology, Volume 1: Immunochemistry, Weir et al. (eds.), Blackwell Scientific Publications, Boston, MA. Prostate cells are typically obtained from a body fluid sample, including blood, semen and urine.

In one aspect of this embodiment of the present invention, cells in a body fluid sample are first contacted with an antibody specific to a prostate cell-specific marker and with an antibody specific for flt-4. These "first" antibodies can be labeled either directly or indirectly with, e.g., a fluorescent marker or a biotin. Alternatively, these "first" antibodies can be reacted with a "second" antibody which is specific for the first antibody, and which second antibody is labeled either directly or indirectly with, e.g., a fluorescent marker. In another aspect, the "first" antibody, which is specific for PSMA, is directly labeled with biotin and the "second" antibody, which is specific for biotin, is directly labeled with a fluorescent marker; the "first" antibody, which is specific for flt-4, is indirectly labeled with a fluorescent marker. The antibody-contacted cells are then assayed in a flow cytometer.

The fluorescent label associated with the prostate cells and the fluorescent label associated with the flt-4 need to fluoresce at different wavelengths, such that the prostate cells and the prostate cells expressing flt-4 can be distinguished. The fluorescence detected by the flow cytometer at the respective different wavelengths allows for the detection of flt-4-expressing prostate cells in the sample by the characteristic profile of forward and side scatter of the cells based on their fluorescence.

In one aspect of this embodiment of the invention, when the body fluid is semen, the cells from such semen sample can be stained with a fluorescent dye specific for DNA, such that the haploid sperm cells can be distinguished from the diploid prostate cells. The haploid cells can then be removed from analysis by specific gating, i.e., DNA signal histogram. However, DNA staining of the cell sample is not always necessary and may depend on the sample in terms of the number of cells and the quality of the sample, such as the number of cells present.

Other detection and separation techniques besides flow cytometry can also provide for the detection of flt-4-expressing prostate cells in a fast manner. One such method is biotin-avidin based separation by affinity chromatography. Typically, such a

technique is performed by incubating the sample of cells with biotin-conjugated antibodies to specific markers, such as PSMA and flt-4, followed by passage through an avidin column. Biotin-antibody-cell complexes bind to the column via the biotin-avidin interaction, while other cells pass through the column. The specificity of the biotin-avidin system is well suited for rapid positive detection and separation.

In another preferred embodiment of the present invention, flt-4 expressing prostate cells can be detected using a laser scanning cytometer. Typically, laser scanning cytometry is performed as follows. A sample containing the cells of interest, *e.g.*, prostate gland biopsy or lymph node biopsy, is fixed on a slide and the cells are contacted with fluorochrome-conjugated antibodies, which allows for the binding of the antibodies to the prostate cell-specific marker and flt-4. The bound cells are then washed and the slide examined using a laser scanning cytometer, which allows for the identification of prostate cells, flt-4 expressing cells and prostate cells expressing flt-4 based on the different fluorescence characteristics imparted by the cell-bound fluorochrome.

Yet another method is magnetic separation using antibody-coated magnetic beads. Kemmner et al., 1992, J. Immunol. Methods 147:197-200; Racila et al., 1998, Proc. Natl. Acad. Sci. USA 95:4589-4594.

In an alternative embodiment, prostate cells expressing flt-4 can be detected by contacting prostate cells with an antibody specific for a flt-4:VEGF-C/D complex. In particular, such an immunoassay is carried out by a method comprising contacting a sample derived from a patient containing prostate cells with an anti-flt-4:VEGF-C/D complex antibody under conditions such that immunospecific binding can occur, and detecting or measuring the amount of any immunospecific binding by the antibody.

The immunoassays which can be used include but are not limited to competitive and non-competitive assay systems using techniques such as Western blots, radioimmunoassays, ELISA (enzyme linked immunosorbent assay), "sandwich" immunoassays, immunoprecipitation assays, precipitin reactions, gel diffusion precipitin reactions, immunodiffusion assays, agglutination assays, complement-fixation assays, immunoradiometric assays, fluorescent immunoassays, protein A immunoassays, to name but a few known in the art.

Kits for diagnostic use are also provided in the present invention, that comprise in one or more containers an anti-flt-4:VEGF-C/D complex antibody or an anti-flt-4 antibody, or an antibody directed to a prostate cell-specific marker, and, optionally, a labeled binding partner to the antibody. Alternatively, the antibody can be labeled with a detectable marker, *e.g.*, a chemiluminescent, enzymatic, fluorescent, or radioactive moiety. A kit is also provided that comprises in one or more containers a nucleic acid probe capable

of hybridizing to *flt-4* and/or a prostate cell-specific marker encoding nucleic acids, *e.g.*, PSMA mRNA. In a specific embodiment, a kit can comprise in one or more containers a pair of primers (*e.g.*, each in the size range of 6-30 nucleotides) that are capable of priming amplification [*e.g.*, by polymerase chain reaction (see, *e.g.*, Innis et al., 1990, PCR Protocols, Academic Press, Inc., San Diego, CA), ligase chain reaction (EP 320,308) use of Q β replicase, cyclic probe reaction, or other methods known in the art] under appropriate reaction conditions of at least a portion of a *flt-4* and/or a prostate cell-specific marker encoding nucleic acids. A kit can optionally further comprise in a container a predetermined amount of a purified *flt-4*:VEGF-C/D complex, or *flt-4* and/or the prostate cell-specific marker or encoding nucleic acids thereof, *e.g.*, for use as a standard or control.

5.2. THERAPEUTICS AND USES THEREOF

The present invention also encompasses a method for treatment, inhibition or prevention of secondary prostate tumor metastases by administration of a therapeutic molecule or composition (termed herein "Therapeutic") which molecule or composition inhibits the expression of the *flt-4* gene product or inhibits a function of wild-type *flt-4*, *e.g.*, *flt-4* binding to VEGF-C/D (*flt-4*:VEGF-C/D complex formation) or tyrosine kinase activity. Such "Therapeutics" include, but are not limited to, the *flt-4* protein, and analogs and derivatives (including fragments) thereof (as described *infra*); antibodies thereto (as described *infra*); nucleic acids encoding *flt-4*, and analogs or derivatives thereof (*e.g.*, as described *infra*); *flt-4* antisense nucleic acids, and agents that inhibit *flt-4* expression or activity, *e.g.*, *flt-4* binding to VEGF-C/D and/or tyrosine kinase activity (*i.e.*, antagonists). In a specific embodiment, the antagonist of *flt-4* activity is a peptidomimetic or peptide analog or organic molecule that binds to *flt-4*. Such an antagonist can be identified by binding assays selected from among those known in the art.

Not intending to be limited to a particular mechanism of action, the surprising discovery that forms, in part, a basis of the present invention is that although it was known that VEGF-C and its binding receptor *flt-4* are implicated in the promotion of lymphoangiogenesis, it was not known that VEGF-C and *flt-4* are involved in an autocrine feedback loop resulting in the promotion and formation of secondary prostate tumor metastases. Further, the inventor discovered that the expression of *flt-4* in a prostate cell indicates that such prostate cell is a cancerous prostate cell that has metastatic potential or is a secondary tumor metastasis of a primary prostate tumor, or is derived therefrom. Thus, in accordance with the present invention, secondary prostate tumors are treated or their growth inhibited or prevented by administration of a Therapeutic that inhibits *flt-4* expression or *flt-4* activity, *e.g.*, *flt-4*:VEGF-C/D complex formation. In a specific embodiment, the

expression of flt-4 is inhibited by administration of an antibody specific for the extracellular domain of flt-4 or by the administration of a nucleic acid comprising a nucleotide sequence complementary to a nucleotide sequence encoding a flt-4 protein. In another specific embodiment, flt-4 function is inhibited by administration of a flt-4 protein or fragment
5 comprising the VEGF-C/D binding region, or a nucleic acid encoding said protein or fragment, which protein or fragment acts as a competitive inhibitor of VEGF-C/D-flt-4 binding.

Therapeutics that antagonize (*i.e.*, reduce or inhibit) flt-4 expression or flt-4 activity include, but are not limited to, flt-4 or an analog, derivative or fragment of flt-4;
10 anti-flt-4:VEGF-C/D complex antibodies (*e.g.*, antibodies specific for a flt-4:VEGF-C/D complex, or a fragment or derivative of the antibody containing the binding region thereof; anti-flt-4 antibodies, or a fragment or derivative of such antibody containing the binding region thereof; nucleic acids encoding flt-4; flt-4 antisense nucleic acids; and flt-4 nucleic acids that are dysfunctional due to, *e.g.*, a heterologous (non-flt-4) insertion within the flt-4
15 coding sequence, that are used to "knockout" endogenous flt-4 function by homologous recombination, see, *e.g.*, Capecchi, 1989, Science 244:1288-1292.

In another embodiment, a Therapeutic of the present invention is a cancer vaccine. Such vaccines include, but are not limited to, dendritic cells or activated T cells. Dendritic cells can be obtained from human donors and once exposed to flt-4 antigen or
20 antigenic fragment, are administered to a prostate cancer patient to activate relevant T cell responses *in vivo*. Alternatively, the dendritic cells are exposed to flt-4 antigen or antigenic fragment *in vitro* and incubated with primed or unprimed T cells to activate the relevant T cell responses *in vitro*. The activated T cells are then administered to a prostate cancer patient. In either alternative, dendritic cells are used to elicit an immunotherapeutic growth
25 inhibiting response against a metastatic prostate tumor.

In a specific embodiment of the present invention, a nucleic acid containing a portion of a flt-4 gene in which flt-4 sequences flank (are both 5' and 3' to) a different gene sequence, is used as a flt-4 antagonist, or to promote flt-4 inactivation by homologous recombination (see also, Koller and Smithies, 1989, Proc. Natl. Acad. Sci. USA 86:8932-
30 8935; Zijlstra et al., 1989, Nature 342: 435-438). Additionally, mutants or derivatives of flt-4 that have greater affinity for VEGF-C/D than wild type flt-4 may be administered to compete with wild type flt-4 protein for VEGF-C/D binding, thereby reducing the levels of VEGF-C/D complexes with wild type flt-4, which results in the inhibition of the autocrine feedback loop. Other Therapeutics that inhibit flt-4 activity, *e.g.*, flt-4:VEGF-C/D complex
35 function can be identified by use of known convenient *in vitro* assays, *e.g.*, based on their

ability to inhibit flt-4:VEGF-C/D binding (complex formation), or as described in Section 5.3, *infra*.

5 In specific embodiments, Therapeutics that antagonize flt-4 expression or activity are administered therapeutically, including prophylactically, in the treatment or inhibition of prostate cancer. A more specific embodiment of the present invention is directed to a method of reducing flt-4 expression or flt-4 activity by targeting mRNAs that express the flt-4 protein. RNA therapeutics currently fall within three classes, antisense species, ribozymes, or RNA aptamers (Good et al., 1997, Gene Therapy 4:45-54).

10 Antisense oligonucleotides have been the most widely used. By way of example, but not limitation, antisense oligonucleotide methodology to reduce flt-4 expression is presented below in Subsection 5.2.2, *infra*. Ribozyme therapy involves the administration, induced expression, etc. of small RNA molecules with enzymatic ability to cleave, bind, or otherwise inactivate specific RNAs, to reduce or eliminate expression of particular proteins (Grassi and Marini, 1996, Annals of Medicine 28:499-510; Gibson,
15 1996, Cancer and Metastasis Reviews 15:287-299). At present, the design of "hairpin" and "hammerhead" RNA ribozymes is necessary to specifically target a particular mRNA such as that for flt-4. RNA aptamers are specific RNA ligand proteins, such as for Tat and Rev RNA (Good et al., 1997, Gene Therapy 4:45-54) that can specifically inhibit their translation. Aptamers specific for flt-4 can be identified by many methods well known in
20 the art, for example, by affecting the formation of a complex in the protein-protein interaction assay described in Section 5.4.1, *infra*.

In another embodiment, the activity or levels of flt-4 are reduced by administration of an antibody that immunospecifically binds to flt-4, or a fragment or a derivative of the antibody containing the binding domain thereof. Such antibodies are
25 described in Section 5.2.3, *infra*. In a preferred embodiment, the anti-flt-4 antibody is obtained from Santa Cruz Corp. (Santa Cruz, CA).

Generally, administration of products of species origin or species reactivity (in the case of antibodies) that is the same species as that of the patient is preferred. Thus, in a preferred embodiment, a human flt-4 protein, or derivative, homolog or analog thereof;
30 nucleic acids encoding human flt-4 or a derivative, homolog or analog thereof; an antibody to a human flt-4 or to a human flt-4:VEGF-C/D complex, or a derivative thereof; or other human agents that affect flt-4 expression or activity, are therapeutically or prophylactically administered to a human patient.

Preferably, suitable *in vitro* or *in vivo* assays are utilized to determine the
35 effect of a specific Therapeutic and whether its administration is indicated for treatment of the affected tissue or individual.

In various specific embodiments, *in vitro* assays can be carried out with representative cells of cell types involved in a patient's disorder, to determine if a Therapeutic has a desired effect upon such cell types.

Therapeutics for use in therapy can be tested in suitable animal model systems prior to testing in humans, including, but not limited to, rats, mice, chicken, cows, monkeys, rabbits, etc. For *in vivo* testing, prior to administration to humans, any animal model system known in the art may be used, including those described in Section 5.6. Illustrative non-limiting examples of models of prostate cancer include those rodent models, human xenograft models, transgenic and reconstitution models, and those spontaneous prostate carcinoma models in dogs and primates listed in Waters et al., 1998, The Prostate 36:47-48; Lucia et al., 1998, The Prostate 36:49-55; Stearns et al., 1998, The Prostate 36:56-58; Green et al., 1998, The Prostate 36:59-63; and Waters et al., 1998, The Prostate 36:64-67, respectively.

5.2.1. GENE THERAPY

In a specific embodiment of the present invention, nucleic acids comprising a sequence encoding flt-4, or a functional derivative thereof, are administered to inhibit flt-4 activity, e.g., flt-4:VEGF-C/D complex activity or formation by way of gene therapy. In more specific embodiments, a nucleic acid encoding the extracellular domain of flt-4 is administered by way of gene therapy. Gene therapy refers to therapy performed by the administration of a nucleic acid to a subject. In this embodiment of the present invention, the nucleic acid expresses its encoded protein(s) that mediates a therapeutic effect by modulating flt-4 activity by interfering with flt-4:VEGF-C/D complex activity or formation. Any of the methods for gene therapy available in the art can be used according to the present invention. Exemplary methods are described below.

For general reviews of the methods of gene therapy, see Goldspiel et al., 1993, Clinical Pharmacy 12:488-505; Wu and Wu, 1991, Biotherapy 3:87-95; Tolstoshev, 1993, Ann. Rev. Pharmacol. Toxicol. 32:573-596; Mulligan, 1993, Science 260:926-932; Morgan and Anderson, 1993, Ann. Rev. Biochem. 62:191-217; and May, 1993, TIBTECH 11:155-215. Methods commonly known in the art of recombinant DNA technology which can be used are described in Ausubel et al., eds., 1993, Current Protocols in Molecular Biology, John Wiley & Sons, NY; and Kriegler, 1990, Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY.

In a preferred aspect, the Therapeutic comprises a flt-4 nucleic acid that is part of an expression vector that expresses a fragment of a flt-4 protein lacking at least a part of the intracellular domain in a suitable host such that the flt-4 fragment no longer has

tyrosine kinase activity but still is able to bind to VEGF-C or VEGF-D. Such a fragment of flt-4 is unable to transduce the intracellular signal which normally occurs upon ligand binding, and thus, binding of VEGF-C or VEGF-D to this fragment of flt-4 will not result in the promotion of angiogenesis and autocrine proliferation of prostate cancer. In particular, such a nucleic acid has a promoter operably linked to the flt-4 coding region, said promoter being inducible or constitutive, and optionally, tissue-specific. In another particular embodiment, a nucleic acid molecule is used in which the flt-4 coding sequence, and any other desired sequences, are flanked by regions that promote homologous recombination at a desired site in the genome, thus providing for intra-chromosomal expression of the flt-4 fragment encoding nucleic acid (Koller and Smithies, 1989, Proc. Natl. Acad. Sci. USA 86:8932-8935; Zijlstra et al., 1989, Nature 342:435-438). In another preferred aspect, the nucleic acid is an antisense nucleic acid that inhibits the expression of flt-4.

Delivery of the nucleic acid into a patient may be either direct, in which case the patient is directly exposed to the nucleic acid or nucleic acid-carrying vector, or indirect, in which case, cells are first transformed with the nucleic acid *in vitro*, then transplanted into the patient. These two approaches are known, respectively, as *in vivo* or *ex vivo* gene therapy.

In a specific embodiment, the nucleic acid is directly administered *in vivo*, where it is expressed to produce the encoded product. This can be accomplished by any of numerous methods known in the art, *e.g.*, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, *e.g.*, by infection using a defective or attenuated retroviral or other viral vector (U.S. Patent No. 4,980,286), or by direct injection of naked DNA, or by use of microparticle bombardment (*e.g.*, a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors, or through use of transfecting agents, by encapsulation in liposomes, microparticles, or microcapsules, or by administering it in linkage to a peptide that is known to enter the nucleus, or by administering it in linkage to a ligand subject to receptor-mediated endocytosis that can be used to target cell types specifically expressing the receptors (*e.g.*, Wu and Wu, 1987, J. Biol. Chem. 262:4429-4432), etc. In another embodiment, a nucleic acid-ligand complex can be formed in which the ligand comprises a fusogenic viral peptide that disrupts endosomes, allowing the nucleic acid to avoid lysosomal degradation. In yet another embodiment, the nucleic acid can be targeted *in vivo* for cell specific uptake and expression, by targeting a specific receptor (see, *e.g.*, International Patent Publications WO 92/06180; WO 92/22635; WO 92/20316; WO 93/14188; and WO 93/20221. Alternatively, the nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression,

by homologous recombination (Koller and Smithies, 1989, Proc. Natl. Acad. Sci. USA 86:8932-8935; Zijlstra et al., 1989, Nature 342:435-438).

In a specific embodiment, a viral vector that contains the *flt-4* encoding nucleic acid is used. In another specific embodiment, a viral vector that contains a *flt-4* antisense nucleic acid is used. For example, a retroviral vector can be used (Miller et al., 1993, Meth. Enzymol. 217:581-599). These retroviral vectors have been modified to delete retroviral sequences that are not necessary for packaging of the viral genome and integration into host cell DNA. The *flt-4* encoding or antisense nucleic acids to be used in gene therapy is/are cloned into the vector, which facilitates delivery of the gene into a patient. More detail about retroviral vectors can be found in Boesen et al., 1994, Biotherapy 6:291-302, which describes the use of a retroviral vector to deliver the *mdr1* gene to hematopoietic stem cells in order to make the stem cells more resistant to chemotherapy. Other references illustrating the use of retroviral vectors in gene therapy are Clowes et al., 1994, J. Clin. Invest. 93:644-651; Kiem et al., 1994, Blood 83:1467-1473; Salmons and Gunzberg, 1993, Human Gene Therapy 4:129-141; and Grossman and Wilson, 1993, Curr. Opin. in Genetics and Devel. 3:110-114.

Adenoviruses are other viral vectors that can be used in gene therapy. Adenoviruses are especially attractive vehicles for delivering genes to respiratory epithelia. Adenoviruses naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are the liver, the central nervous system, endothelial cells (such as prostate cells) and muscle. Adenoviruses have the advantage of being capable of infecting non-dividing cells. Kozarsky and Wilson, 1993, Current Opinion in Genetics and Development 3:499-503, discuss adenovirus-based gene therapy. The use of adenovirus vectors to transfer genes to the respiratory epithelia of rhesus monkeys has been demonstrated by Bout et al., 1994, Human Gene Therapy 5:3-10. Other instances of the use of adenoviruses in gene therapy can be found in Rosenfeld et al., 1991, Science 252:431-434; Rosenfeld et al., 1992, Cell 68:143-155; and Mastrangeli et al., 1993, J. Clin. Invest. 91:225-234.

Adeno-associated virus (AAV) has also been proposed for use in gene therapy (Walsh et al., 1993, Proc. Soc. Exp. Biol. Med. 204:289-300).

Another approach to gene therapy involves transferring a gene into cells in tissue culture by methods such as electroporation, lipofection, calcium phosphate-mediated transfection, or viral infection. Usually, the method of transfer includes the transfer of a selectable marker to the cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene from those that have not. Those cells are then delivered to a patient.

In this embodiment, the nucleic acid is introduced into a cell prior to administration *in vivo* of the resulting recombinant cell. Such introduction can be carried out by any method known in the art including, but not limited to, transfection by electroporation, microinjection, infection with a viral or bacteriophage vector containing the nucleic acid sequences, cell fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion, etc. Numerous techniques are known in the art for the introduction of foreign genes into cells (see, *e.g.*, Loeffler and Behr, 1993, *Meth. Enzymol.* 217:599-618; Cohen et al., 1993, *Meth. Enzymol.* 217:618-644; Cline, 1985, *Pharmac. Ther.* 29:69-92) and may be used in accordance with the present invention, provided that the necessary developmental and physiological functions of the recipient cells are not disrupted. The technique should provide for the stable transfer of the nucleic acid to the cell, so that the nucleic acid is expressible by the cell and preferably, is heritable and expressible by its cell progeny.

The resulting recombinant cells can be delivered to a patient by various methods known in the art. In a preferred embodiment, epithelial cells are injected, *e.g.*, subcutaneously. Recombinant blood cells (*e.g.*, hematopoietic stem or progenitor cells) are preferably administered intravenously. The amount of cells envisioned for use depends on the desired effect, patient state, etc., and can be determined by one skilled in the art.

Cells into which a nucleic acid can be introduced for purposes of gene therapy encompass any desired, available cell type, and include but are not limited to epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes, blood cells such as T lymphocytes, B lymphocytes, monocytes, macrophages, neutrophils, eosinophils, megakaryocytes, and granulocytes, various stem or progenitor cells, in particular hematopoietic stem or progenitor cells, *e.g.*, as obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, etc.

In a preferred embodiment, the cell used for gene therapy is autologous to the patient.

In one embodiment in which recombinant cells are used in gene therapy, a flt-4 fragment encoding nucleic acid is introduced into the cells such that the gene is expressible by the cells or their progeny, and the recombinant cells are then administered *in vivo* for therapeutic effect. In a specific embodiment, stem or progenitor cells are used. Any stem and/or progenitor cells which can be isolated and maintained *in vitro* can potentially be used in accordance with this embodiment of the present invention. Such stem cells include but are not limited to hematopoietic stem cells (HSCs), stem cells of epithelial tissues such as the skin and the lining of the gut, embryonic heart muscle cells, liver stem

cells (International Patent Publication WO 94/08598), and neural stem cells (Stemple and Anderson, 1992, Cell 71:973-985).

Epithelial stem cells (ESCs), or keratinocytes, can be obtained from tissues such as the skin and the lining of the gut by known procedures (Rheinwald, 1980, Meth. Cell Biol. 2A:229). In stratified epithelial tissue such as the skin, renewal occurs by mitosis of stem cells within the germinal layer, the layer closest to the basal lamina. Similarly, stem cells within the lining of the gut provide for a rapid renewal rate of this tissue. ESCs or keratinocytes obtained from the skin or lining of the gut of a patient or donor can be grown in tissue culture (Rheinwald, 1980, Meth. Cell Bio. 2A:229; Pittelkow and Scott, 1986, Mayo Clinic Proc. 61:771). If the ESCs are provided by a donor, a method for suppression of host versus graft reactivity (*e.g.*, irradiation, or drug or antibody administration to promote moderate immunosuppression) can also be used.

With respect to hematopoietic stem cells (HSCs), any technique that provides for the isolation, propagation, and maintenance *in vitro* of HSCs can be used in this embodiment of the invention. Techniques by which this may be accomplished include (a) the isolation and establishment of HSC cultures from bone marrow cells isolated from the future host, or a donor, or (b) the use of previously established long-term HSC cultures, which may be allogeneic or xenogeneic. Non-autologous HSCs are used preferably in conjunction with a method of suppressing transplantation immune reactions between the future host and patient. In a particular embodiment of the present invention, human bone marrow cells can be obtained from the posterior iliac crest by needle aspiration (see, *e.g.*, Kodo et al., 1984, J. Clin. Invest. 73: 1377-1384). In a preferred embodiment of the present invention, the HSCs can be made highly enriched or in substantially pure form. This enrichment can be accomplished before, during, or after long-term culturing, and can be done by any technique known in the art. Long-term cultures of bone marrow cells can be established and maintained by using, for example, modified Dexter cell culture techniques (Dexter et al., 1977, J. Cell Physiol. 91:335) or Witlock-Witte culture techniques (Witlock and Witte, 1982, Proc. Natl. Acad. Sci. USA 79:3608-3612).

In a specific embodiment, the nucleic acid to be introduced for purposes of gene therapy comprises an inducible promoter operably linked to the coding region, such that expression of the nucleic acid is controllable by controlling the presence or absence of the appropriate inducer of transcription.

Additional methods can be adapted for use to deliver a nucleic acid encoding a fragment of flt-4 that binds to VEGF-C/D but lacks tyrosine kinase activity.

5.2.2. USE OF ANTISENSE OLIGONUCLEOTIDES FOR SUPPRESSION OF FLT-4 EXPRESSION

In a specific embodiment of the present invention, flt-4 expression or activity is inhibited by use of flt-4 antisense nucleic acids. The present invention provides the therapeutic or prophylactic use of nucleic acids of at least six nucleotides that are antisense to a gene or cDNA encoding flt-4, or a portion thereof. A flt "antisense" nucleic acid as used herein refers to a nucleic acid capable of hybridizing to a portion of a flt-4 RNA (preferably mRNA) by virtue of some sequence complementarity. The antisense nucleic acid may be complementary to a coding and/or noncoding region of a flt-4 mRNA. One illustrative but non-limiting example of a flt-4 antisense nucleic acid comprises the nucleotide sequence 5'-GGCGCCCCGCTGCAT-3' (SEQ ID NO:3). Such antisense nucleic acids that inhibit expression of flt-4 have utility as Therapeutics, and can be used in the treatment, inhibition or prevention of prostate cancer as described *supra*.

The antisense nucleic acids of the invention can be oligonucleotides that are double-stranded or single-stranded, RNA or DNA, or a modification or derivative thereof, which can be directly administered to a cell, or which can be produced intracellularly by transcription of exogenous, introduced sequences.

In another embodiment, the present invention is directed to a method for inhibiting the expression of flt-4 nucleic acid sequences, in a prokaryotic or eukaryotic cell, comprising providing the cell with an effective amount of a composition comprising a flt-4 antisense nucleic acid, or a derivative thereof, of the invention.

The flt-4 antisense nucleic acids are of at least six nucleotides and are preferably oligonucleotides, ranging from 6 to about 200 nucleotides. In specific aspects, the oligonucleotide is at least 10 nucleotides, at least 15 nucleotides, at least 100 nucleotides, or at least 200 nucleotides. The oligonucleotides can be DNA or RNA or chimeric mixtures, or derivatives or modified versions thereof, and either single-stranded or double-stranded. The oligonucleotide can be modified at the base moiety, sugar moiety, or phosphate backbone. The oligonucleotide may include other appending groups such as peptides, agents facilitating transport across the cell membrane (see, *e.g.*, Letsinger et al., 1989, Proc. Natl. Acad. Sci. U.S.A. 86:6553-6556; Lemaitre et al., 1987, Proc. Natl. Acad. Sci. 84:648-652; International Patent Publication No. WO 88/09810) or blood-brain barrier (see, *e.g.*, International Patent Publication No. WO 89/10134), hybridization-triggered cleavage agents (see, *e.g.*, Krol et al., 1988, BioTechniques 6:958-976), or intercalating agents (see, *e.g.*, Zon, 1988, Pharm. Res. 5:539-549).

In a preferred aspect of the invention, a flt-4 antisense oligonucleotide is provided, preferably as single-stranded DNA. The oligonucleotide may be modified at any position in its structure with constituents generally known in the art.

The flt-4 antisense oligonucleotides may comprise at least one modified base moiety which is selected from the group including but not limited to 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl)uracil, 5-carboxymethylaminomethyl-2-thio-uridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5N-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methyl-thio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine.

In another embodiment, the oligonucleotide comprises at least one modified sugar moiety selected from the group including, but not limited to, arabinose, 2-fluoroarabinose, xylulose, and hexose.

In yet another embodiment, the oligonucleotide comprises at least one modified phosphate backbone selected from the group consisting of a phosphorothioate, a phosphorodithioate, a phosphoramidothioate, a phosphoramidate, a phosphordiamidate, a methylphosphonate, an alkyl phosphotriester, and a formacetal, or an analog of the foregoing.

In yet another embodiment, the oligonucleotide is a 2-a-anomeric oligonucleotide. An a-anomeric oligonucleotide forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units, the strands run parallel to each other (Gautier et al., 1987, Nucl. Acids Res. 15:6625-6641).

The oligonucleotide may be conjugated to another molecule, e.g., a peptide, hybridization-triggered cross-linking agent, transport agent, hybridization-triggered cleavage agent, etc.

Oligonucleotides of the invention may be synthesized by standard methods known in the art, e.g., by use of an automated DNA synthesizer (such as are commercially available from Biosearch, Applied Biosystems, etc.). As examples, phosphorothioate oligonucleotides may be synthesized by the method of Stein et al. (1988, Nucl. Acids Res. 16:3209), methylphosphonate oligonucleotides can be prepared by use of controlled pore glass polymer supports (Sarin et al., 1988, Proc. Natl. Acad. Sci. U.S.A. 85:7448-7451), etc.

In a specific embodiment, the flt-4 antisense oligonucleotides comprise catalytic RNAs, or ribozymes (see, e.g., International Patent Publication No. WO 90/11364;

Sarver et al., 1990, Science 247:1222-1225). In another embodiment, the oligonucleotide is a 2'-O-methylribonucleotide (Inoue et al., 1987, Nucl. Acids Res. 15:6131-6148), or a chimeric RNA-DNA analog (Inoue et al., 1987, FEBS Lett. 215:327-330).

In an alternative embodiment, the flt-4 antisense nucleic acids of the invention are produced intracellularly by transcription from an exogenous sequence. For example, a vector can be introduced in vivo such that it is taken up by a cell, within which cell the vector or a portion thereof is transcribed, producing an antisense nucleic acid (RNA) of the invention. Such a vector would contain a sequence encoding a flt-4 antisense nucleic acid. Such a vector can remain episomal or become chromosomally integrated, as long as it can be transcribed to produce the desired antisense RNA. Such vectors can be constructed by recombinant DNA technology methods standard in the art. Vectors can be plasmid, viral, or others known in the art to be capable of replication and expression in mammalian cells. Expression of the sequences encoding the flt-4 antisense RNAs can be by any promoter known in the art to act in mammalian, preferably human, cells. Such promoters can be inducible or constitutive. Such promoters include, but are not limited to, the SV40 early promoter region (Bernoist and Chambon, 1981, Nature 290:304-310), the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto et al., 1980, Cell 22:787-797), the herpes thymidine kinase promoter (Wagner et al., 1981, Proc. Natl. Acad. Sci. U.S.A. 78:1441-1445), the regulatory sequences of the metallothionein gene (Brinster et al., 1982, Nature 296:39-42), etc.

The antisense nucleic acids of the invention comprise a sequence complementary to at least a portion of an RNA transcript of a flt-4 gene, preferably a human flt-4 gene. However, absolute complementarity, although preferred, is not required. A sequence "complementary to at least a portion of an RNA," as referred to herein, means a sequence having sufficient complementarity to be able to hybridize with the RNA, forming a stable duplex; in the case of double-stranded flt-4 antisense nucleic acids, a single strand of the duplex DNA may thus be tested, or triplex formation may be assayed. The ability to hybridize will depend on both the degree of complementarity and the length of the antisense nucleic acid. Generally, the longer the hybridizing nucleic acid, the more base mismatches with a flt-4 RNA it may contain and still form a stable duplex (or triplex, as the case may be). One skilled in the art can ascertain a tolerable degree of mismatch by use of standard procedures to determine the melting point of the hybridized complex.

The flt-4 antisense nucleic acid can be used to treat (or prevent) secondary prostate tumor metastases. In a preferred embodiment, a single-stranded flt-4 DNA antisense oligonucleotide is used.

Pharmaceutical compositions of the invention (see Section 5.5, *infra*), comprising an effective amount of a flt-4 antisense nucleic acid in a pharmaceutically acceptable carrier can be administered to a patient having a secondary prostate tumor metastasis.

5 The amount of flt-4 antisense nucleic acid that will be effective in the treatment of a particular disorder or condition will depend on the nature of the disorder or condition, and can be determined by standard clinical techniques. Where possible, it is desirable to determine the antisense cytotoxicity in vitro, and then in useful animal model systems, prior to testing and use in humans.

10 In a specific embodiment, pharmaceutical compositions comprising flt-4 antisense nucleic acids are administered via liposomes, microparticles, or microcapsules. In various embodiments of the invention, it may be useful to use such compositions to achieve sustained release of the flt-4 antisense nucleic acids.

15 5.2.3. ANTIBODIES TO FLT-4

According to the present invention, flt-4 or a fragment, derivative or homolog thereof, or a flt-4:VEGF-C/D complex or a fragment, derivative or homolog thereof, may be used as an immunogen to generate antibodies which immunospecifically bind such immunogen. Such antibodies include, but are not limited to, polyclonal,
20 monoclonal, chimeric, single chain, Fab fragments, and an Fab expression library. In a specific embodiment, antibodies to human flt-4 are produced. In another specific embodiment, antibodies to a complex of human flt-4 and human VEGF-C or VEGF-D are produced. In another embodiment, a complex formed from a fragment of flt-4 and a fragment of VEGF-C or VEGF-D, which fragments contain the protein domain that
25 interacts with the other member of the complex, are used as an immunogen for antibody production. One illustrative but non-limiting example of an anti-flt-4 antibody is the anti-flt-4 antibody obtained from Santa Cruz Corp. (Santa Cruz, CA).

In another specific embodiment, the antibody to a flt-4 protein is a bispecific antibody (see generally, *e.g.* Fanger and Drakeman, 1995, Drug News and Perspectives 8:
30 133-137). Such a bispecific antibody is genetically engineered to recognize both (1) a flt-4 epitope and (2) one of a variety of "trigger" molecules, *e.g.* Fc receptors on myeloid cells, and CD3 and CD2 on T cells, that have been identified as being able to cause a cytotoxic T-cell to destroy a particular target. Such bispecific antibodies can be prepared either by chemical conjugation, hybridoma, or recombinant molecular biology techniques known to
35 the skilled artisan.

Various procedures known in the art may be used for the production of polyclonal antibodies to a flt-4 protein, or a fragment, derivative, homolog or analog of the protein or for the production of polyclonal antibodies to a complex of flt-4 and VEGF-C/D.

For production of the antibody, various host animals can be immunized by
5 injection with a native flt-4 protein or a flt-4:VEGF-C/D complex or a synthetic version, or a derivative of the foregoing, such as a cross-linked flt-4:VEGF-C/D complex. Such host animals include, but are not limited to, rabbits, mice, rats, etc. Various adjuvants can be used to increase the immunological response, depending on the host species, and include, but are not limited to, Freund's (complete and incomplete), mineral gels such as aluminum
10 hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, dinitrophenol, and potentially useful human adjuvants such as bacille Calmette-Guerin (BCG) and Corynebacterium parvum.

For preparation of monoclonal antibodies directed towards a flt-4 protein or derivative, fragment, homolog or analog thereof, or a flt-4:VEGF-C/D complex, or a
15 derivative, fragment, homolog or analog thereof, any technique that provides for the production of antibody molecules by continuous cell lines in culture may be used. Such techniques include, but are not restricted to, the hybridoma technique originally developed by Kohler and Milstein (1975, Nature 256:495-497), the trioma technique (Gustafsson et al., 1991, Hum. Antibodies Hybridomas 2:26-32), the human B-cell hybridoma technique
20 (Kozbor et al., 1983, Immunology Today 4:72), and the EBV hybridoma technique to produce human monoclonal antibodies (Cole et al., 1985, In: Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). In an additional embodiment of the invention, monoclonal antibodies can be produced in germ-free animals utilizing recent technology described in International Patent Application PCT/US90/02545.

25 According to the present invention, human antibodies may be used and can be obtained by using human hybridomas (Cote et al., 1983, Proc. Natl. Acad. Sci. USA 80:2026-2030) or by transforming human B cells with EBV virus in vitro (Cole et al., 1985, In: Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). In fact, according to the invention, techniques developed for the production of "chimeric
30 antibodies" (Morrison et al., 1984, Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger et al., 1984, Nature 312:604-608; Takeda et al., 1985, Nature 314:452-454) by splicing the genes from a mouse antibody molecule specific for the flt-4 protein together with genes from a human antibody molecule of appropriate biological activity can be used; such antibodies are within the scope of this invention.

35 According to the present invention, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce flt-4 or flt-

4:VEGF-C/D complex-specific antibodies. An additional embodiment of the invention utilizes the techniques described for the construction of Fab expression libraries (Huse et al., 1989, Science 246:1275-1281) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity for flt-4 or for flt-4:VEGF-C/D complexes, derivatives, or analogs thereof. Non-human antibodies can be "humanized" by known methods (e.g., U.S. Patent No. 5,225,539).

Antibody fragments that contain the idiotypes of flt-4 or flt-4:VEGF-C/D complex can be generated by techniques known in the art. For example, such fragments include, but are not limited to, the F(ab')₂ fragment which can be produced by pepsin digestion of the antibody molecule; the Fab' fragment that can be generated by reducing the disulfide bridges of the F(ab')₂ fragment; the Fab fragment that can be generated by treating the antibody molecular with papain and a reducing agent; and Fv fragments.

In the production of antibodies, screening for the desired antibody can be accomplished by techniques known in the art, e.g., ELISA (enzyme-linked immunosorbent assay). To select antibodies specific to a particular domain of the flt-4 protein, or a derivative, homolog, or analog thereof, one may assay generated hybridomas for a product that binds to the fragment of the flt-4 protein, or a derivative, homolog, or analog thereof, that contains such a domain. For selection of an antibody that specifically binds a flt-4:VEGF-C/D complex, or a derivative, homolog, or analog thereof, but which does not specifically bind to the individual proteins of the flt-4:VEGF-C/D complex, or a derivative, homolog, or analog thereof, one can select on the basis of positive binding to the flt-4:VEGF-C/D complex and a lack of binding to the individual flt-4 and VEGF-C or VEGF-D proteins.

Antibodies specific to a domain of the flt-4 protein or to a domain of the flt-4:VEGF-C/D complex, or a derivative, homolog, or analog thereof, are also provided.

The foregoing antibodies can be used in methods known in the art relating to the localization and/or quantification of flt-4 or flt-4:VEGF-C/D complexes, e.g., for imaging these proteins, measuring levels thereof in appropriate physiological samples, in diagnostic methods, etc. This hold true also for a derivative, homolog, or analog thereof of a flt-4 protein or of a flt-4:VEGF-C/D complex.

In another embodiment of the invention (see *infra*), an anti-flt-4 antibody or an anti-flt-4:VEGF-C/D complex antibody or a fragment thereof containing the binding domain, is a Therapeutic.

Antibodies and antigen-binding antibody fragments may also be conjugated to a heterologous protein or peptide by chemical conjugation or recombinant DNA technology. The resultant chimeric protein possesses the antigen-binding specificity of the

antibody and the function of the heterologous protein. For example, a polynucleotide encoding the antigen binding region of an antibody specific for the extracellular domain of flt-4 can be genetically fused to a coding sequence for the zeta chain of the T cell receptor. After expressing this construct in T cells, the T cells are expanded *ex vivo* and infused into a prostate cancer patient. T cells expressing this chimeric protein are specifically directed to tumors that express flt-4 as a result of the antibody binding specificity and cause tumor cell killing. Alternatively, an antibody is fused to a protein which induces migration of leukocytes or has an affinity to attract other compounds to a tumor site. A specific protein of this type is streptavidin. The binding of a streptavidin conjugated antibody to a tumor cell can be followed by the addition of a biotinylated drug, toxin or radioisotope to cause tumor specific killing.

Kits for use with such *in vitro* tumor localization and therapy methods containing the monoclonal antibodies (or fragments thereof) conjugated to any of the above types of substances can be prepared. The components of the kits can be packaged either in aqueous medium or in lyophilized form. When the monoclonal antibodies (or fragments thereof) are used in the kits in the form of conjugates in which a label or a therapeutic moiety is attached, such as a radioactive metal ion or a therapeutic drug moiety, the components of such conjugates can be supplied either in fully conjugated form, in the form of intermediates or as separate moieties to be conjugated by the user of the kit.

5.3. ASSAYS OF FLT-4:VEGF-C/D COMPLEXES AND DERIVATIVES AND ANALOGS THEREOF

The functional activity of flt-4, as measured by its ability to form a flt-4:VEGF-C/D complex, or a derivative, fragment or analog thereof, can be assayed by various methods. Potential antagonists of flt-4:VEGF-C/D complex formation, *e.g.*, anti-flt-4:VEGF-C/D complex antibodies and flt-4 antisense nucleic acids, can be assayed for the ability to inhibit flt-4:VEGF-C/D complex formation.

In one embodiment of the present invention, where one is assaying for the ability to bind or compete with a wild-type flt-4:VEGF-C/D complex for binding to an anti-flt-4:VEGF-C/D complex antibody, various immunoassays known in the art can be used, including but not limited to competitive and non-competitive assay systems using techniques such as radioimmunoassay, ELISA (enzyme linked immunosorbent assay), "sandwich" immunoassays, immunoradiometric assays, gel diffusion precipitin reactions, immunodiffusion assays, in situ immunoassays (using colloidal gold, enzyme or radioisotope labels), western blot analysis, precipitation reactions, agglutination assays (*e.g.*, gel agglutination assays, hemagglutination assays), complement fixation assays, immunofluorescence assays, protein A assays, immunoelectrophoresis assays, etc. In one

embodiment, antibody binding is detected by detecting a label on the primary antibody. In another embodiment, the primary antibody is detected by detecting binding of a secondary antibody or reagent to the primary antibody. In a further embodiment, the secondary antibody is labeled. Many means are known in the art for detecting binding in an immunoassay and are within the scope of the present invention.

The expression of the *flt-4* gene (both endogenous and those expressed from cloned DNA containing the genes) can be detected using techniques known in the art, including but not limited to Southern hybridization (Southern, 1975, J. Mol. Biol. 98:503-517), northern hybridization (see, *e.g.*, Freeman et al., 1983, Proc. Natl. Acad. Sci. USA 80:4094-4098), restriction endonuclease mapping (Sambrook et al., 1989, Molecular Cloning, A Laboratory Manual, 2nd Ed. Cold Spring Harbor Laboratory Press, New York), RNase protection assays (Current Protocols in Molecular Biology, John Wiley and Sons, New York, 1997), DNA sequence analysis, and polymerase chain reaction amplification (PCR; U.S. Patent Nos. 4,683,202, 4,683,195, and 4,889,818; Gyllenstein et al., 1988, Proc. Natl. Acad. Sci. USA 85:7652-7657; Ochman et al., 1988, Genetics 120:621-623; Loh et al., 1989, Science 243:217-220) followed by Southern hybridization with probes specific for the *flt-4* gene, in various cell types. Methods of amplification other than PCR commonly known in the art can be employed. The stringency of the hybridization conditions for northern or Southern blot analysis can be manipulated to ensure detection of nucleic acids with the desired degree of relatedness to the specific probes used. Modifications to these methods and other methods commonly known in the art can be used.

Derivatives (*e.g.*, fragments), homologs and analogs of *flt-4* can be assayed for binding to VEGF-C/D by any method known in the art, for example the modified yeast matrix mating test described in Section 5.4.1, *infra*; immunoprecipitation with an antibody that binds to *flt-4* complexed with other proteins, followed by size fractionation of the immunoprecipitated proteins (*e.g.*, by denaturing or nondenaturing polyacrylamide gel electrophoresis); Western blot analysis, etc.

One embodiment of the invention provides a method for screening a derivative, homolog or analog of *flt-4* for biological activity comprising contacting said derivative, homolog or analog of *flt-4* with VEGF-C/D; and detecting the formation of a complex between said derivative, homolog or analog of *flt-4* and VEGF-C/D; wherein detecting formation of said complex indicates that said derivative, homolog or analog of *flt-4* has biological (*e.g.*, binding) activity.

5.4. SCREENING FOR ANTAGONISTS OF FLT-4 ACTIVITY

A functional activity of flt-4 is its ability to bind its ligand, VEGF-C or VEGF-D. Thus, flt-4:VEGF-C/D complexes, and derivatives, fragments and analogs thereof, nucleic acids encoding flt-4, VEGF-C and VEGF-D as well as derivatives, fragments and analogs of the nucleic acids, can be used to screen for compounds that bind to or modulate the function of, flt-4:VEGF-C/D complex encoding nucleic acids, complex member proteins, and derivatives of the foregoing, and thus, have potential use as agonists or antagonists of flt-4:VEGF-C/D complex activity or formation. The present invention is thus directed to assays for detecting molecules that specifically bind to, or modulate the function of, flt-4, VEGF-C and VEGF-D nucleic acids, proteins or derivatives of the nucleic acids and proteins. For example, recombinant cells expressing both flt-4, VEGF-C and VEGF-D nucleic acids can be used to recombinantly produce the complexes or proteins in these assays, to screen for molecules that bind to, or interfere with, or promote flt-4:VEGF-C/D complex formation or activity. In preferred embodiments, polypeptide analogs that have superior stabilities but retain the ability to form a flt-4:VEGF-C/D complex (*e.g.*, flt-4 and VEGF-C or VEGF-D modified to be resistant to proteolytic degradation in the binding assay buffers, or to be resistant to oxidative degradation), are used to screen for modulators of flt-4 activity or flt-4:VEGF-C/D complex activity or formation. Such resistant molecules can be generated, *e.g.*, by substitution of amino acids at proteolytic cleavage sites, the use of chemically derivatized amino acids at proteolytic susceptible sites, and the replacement of amino acid residues subject to oxidation, *i.e.* methionine and cysteine.

A molecule (*e.g.*, putative binding partner or modulator of flt-4:VEGF-C/D complex activity or formation) is contacted with the flt-4:VEGF-C/D complex, or fragment thereof, under conditions conducive to binding or modulation, and then a molecule that specifically bind to or modulate flt-4:VEGF-C/D complex activity or formation is identified. Similar methods can be used to screen for molecules that bind to or modulate the function of flt-4:VEGF-C/D nucleic acids or derivatives thereof.

A particular aspect of the present invention relates to identifying molecules that inhibit or promote formation or degradation of a flt-4:VEGF-C/D complex, *e.g.*, using the method described for screening inhibitors using the modified yeast matrix mating test described in Section 5.6.1., *infra*, and International Patent Publication WO 97/47763 entitled "Identification and Comparison of Protein-Protein Interactions that Occur in Populations and Identification of Inhibitors of These Interactions", which is incorporated by reference herein in its entirety.

In one embodiment of the invention, a molecule that modulates activity of flt-4 or a complex of flt-4 and VEGF-C/D, is identified by contacting one or more candidate

molecules with flt-4 in the presence of VEGF-C or VEGF-D; and measuring the amount of complex that forms between flt-4 and VEGF-C or VEGF-D; wherein an increase or decrease in the amount of complex that forms relative to the amount that forms in the absence of the candidate molecule(s) indicates that the molecule(s) modulates the activity of flt-4 or VEGF-C or VEGF-D or said complex of flt-4 and VEGF-C or VEGF-D. In preferred embodiments, a modulator is identified by administering a candidate molecule to a transgenic non-human animal expressing both flt-4 and VEGF-C or flt-4 and VEGF-D from promoters that are not the native flt-4 or the native VEGF-C or VEGF-D promoters, more preferably where the candidate molecule is also recombinantly expressed in the transgenic non-human animal. Alternatively, the method for identifying such a modulator can be carried out *in vitro*, preferably with purified flt-4, purified VEGF-C or VEGF-D, and a purified candidate molecule.

Methods that can be used to carry out the foregoing are commonly known in the art. Agents/molecules to be screened can be provided as mixtures of a limited number of specified compounds, or as compound libraries, peptide libraries and the like. Agents/molecules to be screened may also include all forms of antisera, antisense nucleic acids, etc., that can modulate flt-4:VEGF-C/D complex activity or formation.

By way of example, diversity libraries, such as random or combinatorial peptide or non-peptide libraries, can be screened for molecules that specifically bind to a flt-4:VEGF-C/D complex. Many libraries are known in the art that can be used, *e.g.*, chemically-synthesized libraries, recombinant, *e.g.*, phage display, libraries, and *in vitro* translation-based libraries.

Examples of chemically-synthesized libraries are described in Fodor et al., 1991, Science 251:767-773; Houghten et al., 1991, Nature 354:84-86; Lam et al., 1991, Nature 354:82-84; Medynski, 1994, BioTechnology 12:709-710; Gallop et al., 1994, J. Medicinal Chemistry 37:1233-1251; Ohlmeyer et al., 1993, Proc. Natl. Acad. Sci. USA 90:10922-10926; Erb et al., 1994, Proc. Natl. Acad. Sci. USA 91:11422-11426; Houghten et al., 1992, BioTechniques 13:412; Jayawickreme et al., 1994, Proc. Natl. Acad. Sci. USA 91:1614-1618; Salmon et al., 1993, Proc. Natl. Acad. Sci. USA 90:11708-11712; International Patent Publication No. WO 93/20242; and Brenner and Lerner, 1992, Proc. Natl. Acad. Sci. USA 89:5381-5383.

Examples of phage display libraries are described in Scott and Smith, 1990, Science 249:386-390; Devlin et al., 1990, Science, 249:404-406; Christian et al., 1992, J. Mol. Biol. 227:711-718; Lenstra, 1992, J. Immunol. Meth. 152:149-157; Kay et al., 1993, Gene 128:59-65; and International Patent Publication No. WO 94/18318.

In vitro translation-based libraries include but are not limited to those described in International Patent Publication No. WO 91/05058; and Mattheakis et al., 1994, Proc. Natl. Acad. Sci. USA 91:9022-9026.

By way of examples of non-peptide libraries, a benzodiazepine library (*e.g.*,
5 Bunin et al., 1994, Proc. Natl. Acad. Sci. USA 91:4708-4712) can be adapted for use. Peptoid libraries (Simon et al., 1992, Proc. Natl. Acad. Sci. USA 89:9367-9371) can also be used. Another example of a library that can be used, in which the amide functionalities in peptides have been permethylated to generate a chemically-transformed combinatorial library, is described by Ostresh et al. (1994, Proc. Natl. Acad. Sci. USA 91:11138-11142).

10 Screening the libraries can be accomplished by any of a variety of commonly known methods. See, *e.g.*, the following references, which disclose screening of peptide libraries: Parmley and Smith, 1989, Adv. Exp. Med. Biol. 251:215-218; Scott and Smith, 1990, Science 249:386-390; Fowlkes et al., 1992, BioTechniques 13:422-427; Oldenburg et al., 1992, Proc. Natl. Acad. Sci. USA 89:5393-5397; Yu et al., 1994, Cell 76:933-945;
15 Staudt et al., 1988, Science 241:577-580; Bock et al., 1992, Nature 355:564-566; Tuerk et al., 1992, Proc. Natl. Acad. Sci. USA 89:6988-6992; Ellington et al., 1992, Nature 355:850-852; U.S. Patent No. 5,096,815, U.S. Patent No. 5,223,409, and U.S. Patent No. 5,198,346, all to Ladner et al.; Rebar and Pabo, 1993, Science 263:671-673; and International Patent Publication No. WO 94/18318.

20 In a specific embodiment, screening can be carried out by contacting the library members with a flt-4:VEGF-C/D complex (or encoding nucleic acid or derivative) immobilized on a solid phase, and harvesting those library members that bind to the protein (or encoding nucleic acid or derivative). Examples of such screening methods, termed "panning" techniques, are described by way of example in Parmley and Smith, 1988, Gene
25 73:305-318; Fowlkes et al., 1992, BioTechniques 13:422-427; International Patent Publication No. WO 94/18318; and in references cited hereinabove.

In a specific embodiment, fragments and/or analogs of flt-4 or VEGF-C or VEGF-D, especially peptidomimetics, are screened for activity as competitive or non-competitive inhibitors of flt-4:VEGF-C/D complex formation, which thereby inhibit flt-
30 4:VEGF-C/D complex activity or formation.

In a preferred embodiment, molecules that bind to flt-4:VEGF-C/D complexes can be screened for using the modified yeast matrix mating test described in Section 5.4.1, *infra*.

In one embodiment, agents that modulate (*i.e.*, antagonize or agonize) flt-
35 4:VEGF-C/D complex activity or formation (flt-4 activity) can be screened for using a binding inhibition assay, wherein agents are screened for their ability to modulate formation

of a flt-4:VEGF-C/D complex under aqueous, or physiological, binding conditions in which flt-4:VEGF-C/D complex formation occurs in the absence of the agent to be tested. Agents that interfere with the formation of flt-4:VEGF-C/D complexes are identified as antagonists of complex formation. Agents that promote the formation of flt-4:VEGF-C/D complexes are identified as agonists of complex formation. Agents that completely block the formation of flt-4:VEGF-C/D complexes are identified as inhibitors of complex formation.

Methods for screening may involve labeling the complex proteins with radioligands (e.g., ^{125}I or ^3H), magnetic ligands (e.g., paramagnetic beads covalently attached to photobiotin acetate), fluorescent ligands (e.g., fluorescein or rhodamine), or enzyme ligands (e.g., luciferase or beta-galactosidase). The reactants that bind in solution can then be isolated by one of many techniques known in the art, including but not restricted to, co-immunoprecipitation of the labeled complex moiety using antisera against the unlabeled binding partner (or labeled binding partner with a distinguishable marker from that used on the second labeled complex moiety), immunoaffinity chromatography, size exclusion chromatography, and gradient density centrifugation. In a preferred embodiment, the labeled binding partner is a small fragment or peptidomimetic that is not retained by a commercially available filter. Upon binding, the labeled species is then unable to pass through the filter, providing for a simple assay of complex formation.

Methods commonly known in the art are used to label at least one of the members of the flt-4:VEGF-C/D complex. Suitable labeling methods include, but are not limited to, radiolabeling by incorporation of radiolabeled amino acids, e.g., ^3H -leucine or ^{35}S -methionine, radiolabeling by post-translational iodination with ^{125}I or ^{131}I using the chloramine T method, Bolton-Hunter reagents, etc., or labeling with ^{32}P using phosphorylase and inorganic radiolabeled phosphorous, biotin labeling with photobiotin-acetate and sunlamp exposure, etc. In cases where one of the members of the flt-4:VEGF-C/D complex is immobilized, e.g., as described *infra*, the free species is labeled. Where neither of the interacting species is immobilized, each can be labeled with a distinguishable marker such that isolation of both moieties can be followed to provide for more accurate quantification, and to distinguish the formation of homomeric from heteromeric complexes. Methods that utilize accessory proteins that bind to one of the modified interactants to improve the sensitivity of detection, increase the stability of the complex, etc. are provided.

Typical binding conditions are, for example, but not by way of limitation, in an aqueous salt solution of 10-250 mM NaCl, 5-50 mM Tris-HCl, pH 5-8, and 0.5% Triton X-100 or other detergent that improves specificity of interaction. Metal chelators and/or divalent cations may be added to improve binding and/or reduce proteolysis. Reaction temperatures may include 4, 10, 15, 22, 25, 35, or 42 degrees Celsius, and time of

incubation is typically at least 15 seconds, but longer times are preferred to allow binding equilibrium to occur. Particular flt-4:VEGF-C/D complexes can be assayed using routine protein binding assays, as described *infra*, to determine optimal binding conditions for reproducible binding.

- 5 The physical parameters of complex formation can be analyzed by quantification of complex formation using assay methods specific for the label used, *e.g.*, liquid scintillation counting for radioactivity detection, enzyme activity for enzyme-labeled moieties etc. The reaction results are then analyzed utilizing Scatchard analysis, Hill analysis, and other methods commonly known in the arts (see, *e.g.*, Proteins, Structures, and
10 Molecular Principles, 2nd Edition (1993) Creighton, Ed., W.H. Freeman and Company, New York).

- In a second common approach to binding assays, one of the binding species is immobilized on a filter, in a microtiter plate well, in a test tube, to a chromatography matrix, etc., either covalently or non-covalently. Proteins can be covalently immobilized
15 using any method well known in the art, for example, but not limited to the method of Kadonaga and Tjian, 1986, Proc. Natl. Acad. Sci. USA 83:5889-5893, *i.e.*, linkage to a cyanogen-bromide derivatized substrate such as CNBr-Sepharose 4B (Pharmacia). Where needed, the use of spacers can reduce steric hindrance by the substrate. Non-covalent attachment of proteins to a substrate include, but are not limited to, attachment of a protein
20 to a charged surface, binding with specific antibodies, binding to a third unrelated interacting protein, etc.

- In one embodiment, immobilized flt-4 is used to assay for binding to radioactively-labeled VEGF-C or VEGF-D in the presence and absence of a compound to be tested for its ability to modulate flt-4:VEGF-C/D complex formation. The binding
25 partners are allowed to bind under aqueous, or physiological, conditions (*e.g.*, the conditions under which the original interaction was detected). Conversely, in another embodiment, VEGF-C or VEGF-D is immobilized and contacted with a labeled flt-4 protein or derivative thereof under binding conditions.

- Assays of agents (including cell extracts or a library pool) for competition
30 for binding of one member of a flt-4:VEGF-C/D complex (or derivatives thereof) with the other member of the flt-4:VEGF-C/D complex labeled by any means (*e.g.*, those means described above) are provided to screen for competitors or enhancers of flt-4:VEGF-C/D complex formation.

- In specific embodiments, blocking agents to inhibit non-specific binding of
35 reagents to other protein components, or absorptive losses of reagents to plastics, immobilization matrices, etc., are included in the assay mixture. Blocking agents include,

but are not restricted to bovine serum albumin, beta-casein, nonfat dried milk, Denhardt's reagent, Ficoll, polyvinylpyrrolidone, nonionic detergents (NP40, Triton X-100, Tween 20, Tween 80, etc.), ionic detergents (e.g., SDS, LDS, etc.), polyethylene glycol, etc.

Appropriate blocking agent concentrations allow flt-4:VEGF-C/D complex formation.

- 5 After binding is performed, unbound, labeled protein is removed in the supernatant, and the immobilized protein retaining any bound, labeled protein is washed extensively. The amount of bound label is then quantified using standard methods in the art to detect the label as described supra.

10 5.4.1. ASSAYS FOR PROTEIN-PROTEIN INTERACTIONS

- One aspect of the present invention provides methods for assaying and screening a fragment, derivative or analog of a flt-4-interacting protein for binding to a flt-4 peptide, or a fragment, derivative, homolog or analog of flt-4. Derivatives, analogs and fragments of VEGF-C or VEGF-D that interact with flt-4 or a derivative, analog or
15 fragment of flt-4 can be identified by means of a yeast matrix mating test system or, more preferably, an improvement thereof as described in International Patent Publication No. WO 97/47763. Because the interactions are screened for in yeast, the intermolecular protein interactions detected in this system occur under physiological conditions that mimic the conditions in mammalian cells (Chien et al., 1991, Proc. Natl. Acad. Sci. USA 88:9578-
20 9581).

- Identification of interacting proteins by the improved yeast matrix mating test is based upon the detection of the expression of a reporter gene ("Reporter Gene"), the transcription of which is dependent upon the reconstitution of a transcriptional regulator by the interaction of two proteins, each fused to one half of the transcriptional regulator. The
25 bait flt-4 or derivative, homolog or analog and prey proteins (proteins to be tested for ability to interact with the bait) are expressed as fusion proteins to a DNA binding domain, and to a transcriptional regulatory domain, respectively, or *vice versa*. In various specific embodiments, the prey has a complexity of at least 50, 100, 500, 1,000, 5,000, 10,000, or 50,000; or has a complexity in the range of 25 to 100,000, 100 to 100,000, 50,000 to
30 100,000, or 100,000 to 500,000. For example, the prey population can be one or more nucleic acids encoding mutants of VEGF-C or VEGF-D, e.g., as generated by site-directed mutagenesis or another method of making mutations in a nucleotide sequence. Preferably, the prey populations are proteins encoded by DNA, e.g., cDNA, genomic DNA, or synthetically-generated DNA. For example, the populations can be expressed from
35 chimeric genes comprising cDNA sequences from an uncharacterized sample of a

population of cDNA from mammalian RNA. Preferably, the prey population are proteins encoded by DNA, *e.g.*, cDNA or genomic DNA or synthetically-generated DNA.

In a specific embodiment, recombinant biological libraries expressing random peptides can be used as the source of prey nucleic acids.

5 In another embodiment of the present invention, the invention provides a method of screening for inhibitors or enhancers of the interacting proteins identified herein. Briefly, the protein-protein interaction assay can be carried out as described herein, except that it is done in the presence of one or more candidate molecules. An increase or decrease in Reporter Gene activity relative to that present when the one or more candidate molecules
10 are absent indicates that the candidate molecule has an effect on the interacting pair. In a preferred method, inhibition of the interaction is selected for (*i.e.*, inhibition of the interaction is necessary for the cells to survive), for example, where the interaction activates the URA3 gene, causing yeast to die in medium containing the chemical 5-fluoroorotic acid (Rothstein, 1983, Meth. Enzymol. 101:167-180). The identification of inhibitors of such
15 interactions can also be accomplished, for example, but not by way of limitation, using competitive inhibitor assays, as described, *supra*.

In general, proteins of the bait and prey populations are provided as fusion (chimeric) proteins (preferably by recombinant expression of a chimeric coding sequence) containing each protein contiguous to a pre-selected sequence. For one population, the pre-
20 selected sequence is a DNA binding domain. The DNA binding domain can be any DNA binding domain, as long as it specifically recognizes a DNA sequence within a promoter, or a DNA sequence that modulates the activity of an DNA promoter, *e.g.*, an enhancer element. For example, the DNA binding domain is of a transcriptional activator or inhibitor. For the other population, the pre-selected sequence is an activator or inhibitor
25 domain of a transcriptional activator or inhibitor, respectively. The regulatory domain alone (not as a fusion to a protein sequence) and the DNA-binding domain alone (not as a fusion to a protein sequence) preferably do not interact detectably so as to avoid false positives in the assay. The assay system further includes a reporter gene operably linked to a promoter that contains a binding site for the DNA binding domain of the transcriptional activator (or
30 inhibitor). Accordingly, in the method of the invention, binding of a flt-4 fusion protein to a prey fusion protein leads to reconstitution of a transcriptional activator (or inhibitor) which activates (or inhibits) expression of the Reporter Gene. The activation of transcription of the Reporter Gene occurs intracellularly, *e.g.*, in prokaryotic or eukaryotic cells, preferably in cell culture.

35 The promoter that is operably linked to the reporter gene nucleotide sequence can be a native or non-native promoter of the nucleotide sequence, and the DNA

binding site(s) that are recognized by the DNA binding domain portion of the fusion protein can be native to the promoter (if the promoter normally contains such binding site(s)) or non-native. Thus, for example, one or more tandem copies (*e.g.*, 4 or 5 copies) of the appropriate DNA binding site can be introduced upstream of the TATA box in the desired promoter (*e.g.*, in the area of position -100 to -400). In a preferred aspect, 4 or 5 tandem copies of the 17 bp UAS (GAL4 DNA binding site) are introduced upstream of the TATA box in the desired promoter, which is upstream of the desired coding sequence for a selectable or detectable marker. In a preferred embodiment, the GAL1-10 promoter is operably fused to the desired nucleotide sequence; the GAL1-10 promoter already contains 5 binding sites for GAL4. Alternatively, the transcriptional activation binding site of the desired gene(s) can be deleted and replaced with GAL4 binding sites (Bartel et al., 1993, *BioTechniques* 14(6):920-924; Chasman et al., 1989, *Mol. Cell. Biol.* 9:4746-4749). The Reporter Gene preferably contains the sequence encoding a detectable or selectable marker the expression of which is regulated by the transcriptional activator, such that the marker is either turned on or off in the cell in response to the presence of a specific interaction. Preferably, the assay is carried out in the absence of background levels of the transcriptional activator (*e.g.*, in a cell that is mutant or otherwise lacking in the transcriptional activator). In one embodiment, more than one Reporter Gene is used to detect transcriptional activation, *e.g.*, one Reporter Gene encoding a detectable marker and one or more Reporter Genes encoding different selectable markers. The detectable marker can be any molecule that can give rise to a detectable signal, *e.g.*, a fluorescent protein or a protein that can be readily visualized or that is recognizable by a specific antibody. The selectable marker can be any protein molecule that confers ability to grow under conditions that do not support the growth of cells not expressing the selectable marker, *e.g.*, the selectable marker is an enzyme that provides an essential nutrient and the cell in which the interaction assay occurs is deficient in the enzyme and the selection medium lacks such nutrient. The Reporter Gene can either be under the control of the native promoter that naturally contains a binding site for the DNA binding protein, or under the control of a heterologous or synthetic promoter.

The activation domain and DNA binding domain used in the assay can be from a wide variety of transcriptional activator proteins, as long as these transcriptional activators have separable binding and transcriptional activation domains. For example, the Gal4 protein of *S. cerevisiae*, the Gcn4 protein of *S. cerevisiae* (Hope and Struhl, 1986, *Cell* 46:885-894), the Ard1 protein of *S. cerevisiae* (Thukral et al., 1989, *Mol. Cell. Biol.* 9:2360-2369), Ace1 regulatory protein of *S. cerevisiae* (Thiele et al., 1988, *Mol. Cell. Biol.* 8: 2745-2752), LexA repressor protein of *E. coli* (Schnarr et al., 1991, *Biochimie* 73:423-431), and herpesvirus VP16 transactivator (Hippenmeyer et al., 1995, *Curr. Opin.*

Biotechnol. 6: 548-552), and the human estrogen receptor (Kumar et al., 1987, Cell 51:941-951) have separable DNA binding and activation domains. The DNA binding domain and activation domain that are employed in the fusion proteins need not be from the same transcriptional activator. In a specific embodiment, a Gal4 or LexA DNA binding domain is employed. In another specific embodiment, a Gal4 or herpes simplex virus VP16 (Triezenberg et al., 1988, Genes Dev. 2:730-742) activation domain is employed. In a specific embodiment, amino acids 1-147 of Gal4 (Ma et al., 1987, Cell 48:847-853; Ptashne et al., 1990, Nature 346:329-331) is the DNA binding domain, and amino acids 411-455 of VP16 (Triezenberg et al., 1988, Genes Dev. 2:730-742; Cress et al., 1991, Science 251:87-90) is the activation domain.

In a preferred embodiment, the yeast transcription factor Gal4 is reconstituted by the protein-protein interaction and the host strain is mutant for Gal4. In another embodiment, the DNA-binding domain is Ace1N and/or the activation domain is Ace1N, the DNA binding and activation domains of the Ace1N protein, respectively. Ace1N is a yeast protein that activates transcription from the CUP1 operon in the presence of divalent copper. CUP1 encodes metallothionein, which chelates copper, and the expression of Cup1 protein allows growth in the presence of copper, which is otherwise toxic to the host cells. The Reporter Gene can also be a CUP1-lacZ fusion that expresses the enzyme β -galactosidase (detectable by routine chromogenic assay) upon binding of a reconstituted Ace1 transcriptional activator (Chaudhuri et al., 1995, FEBS Letters 357:221-226). In another specific embodiment, the DNA binding domain of the human estrogen receptor is used, with a Reporter Gene driven by one or three estrogen receptor response elements (Le Douarin et al., 1995, Nucl. Acids. Res. 23:876-878).

The DNA binding domain and the transcription activator/inhibitor domain each preferably has a nuclear localization signal (Ylikomi et al., 1992, EMBO J. 11:3681-3694; Dingwall and Laskey, 1991, Trends Biochem. Sci. 16:479-481) functional in the cell in which the fusion proteins are to be expressed.

To facilitate isolation of the encoded proteins, the fusion constructs can further contain sequences encoding affinity tags such as glutathione-S-transferase or maltose-binding protein or an epitope of an available antibody, for affinity purification (*e.g.*, binding to glutathione, maltose, or a particular antibody specific for the epitope, respectively) (Allen et al., 1995, Trends Biochem. Sci. 20:511-516). In another embodiment, the fusion constructs further comprise bacterial promoter sequences for recombinant production of the fusion protein in bacterial cells (Allen et al., 1995, Trends Biochem. Sci. 20:511-516).

The host cell in which the interaction assay occurs can be any cell, prokaryotic or eukaryotic, in which transcription of the Reporter Gene can occur and be detected, including but not limited to mammalian (*e.g.*, monkey, chicken, mouse, rat, human, bovine), bacterial, and insect cells, and is preferably a yeast cell. Expression constructs encoding and capable of expressing the binding domain fusion proteins, the transcriptional activation domain fusion proteins, and the Reporter Gene product(s), are provided within the host cell, by mating of cells containing the expression constructs, or by cell fusion, transformation, electroporation, microinjection, etc. In a specific embodiment in which the assay is carried out in mammalian cells, the DNA binding domain is the GAL4 DNA binding domain, the activation domain is the herpes simplex virus VP16 transcriptional activation domain, and the Reporter Gene contains the desired reporter gene coding sequence(s) operably linked to a minimal promoter element from the adenovirus E1B gene driven by several GAL4 DNA binding sites (Fearon et al., 1992, Proc. Natl. Acad. Sci. USA 89:7958-7962). The host cell used should not express an endogenous transcription factor that binds to the same DNA site as that recognized by the DNA binding domain fusion population. Also, preferably, the host cell is mutant or otherwise lacking in an endogenous, functional form of the Reporter Gene(s) used in the assay.

Various vectors and host strains for expression of the two fusion protein populations in yeast are known and can be used (see, *e.g.*, Fields et al., U.S. Patent No. 5,146,614; Bartel et al., 1993, "Using the two-hybrid system to detect protein-protein interactions," in Cellular Interactions in Development, Hartley, ed., Practical Approach Series xviii, IRL Press at Oxford University Press, New York, NY, pp. 153-179; Fields and Sternglanz, 1994, Trends in Genetics 10:286-292). By way of example but not limitation, yeast strains or derivative strains made therefrom, which can be used are N105, N106, N1051, N1061, and YULH. Exemplary strains that can be used in the assay of the invention also include, but are not limited to, the following:

Y190: *MATa, ura3-52, his3-200, lys2-801, ade2-101, trp1-901, leu2-3,112, gal4 α , gal80 α , cyh'2, LYS2::GALI_{UAS}-HIS3_{TATA}HIS3, URA3::GALI_{UAS}-GALI_{TATA}-lacZ* (available from Clontech, Palo Alto, CA; Harper et al., 1993, Cell 75:805-816). Y190 contains HIS3 and lacZ Reporter Genes driven by GAL4 binding sites.

CG-1945: *MATa, ura3-52, his3-200, lys2-801, ade2-101, trp1-901, leu2-3,112, gal4-542, gal80-538, cyh'2, LYS2::GALI_{UAS}-HIS3_{TATA}HIS3, URA3::GALI_{UAS17mers(x3)}-CYC1_{TATA}-lacZ* (available from Clontech, Palo Alto, CA). CG-1945 contains HIS3 and lacZ Reporter Genes driven by GAL4 binding sites.

Y187: *MAT α* , *ura3-52*, *his3-200*, *ade2-101*, *trp1-901*, *leu2-3,112*, *gal4 α* , *gal80 α* , *URA3::GAL1_{UAS}-GAL1_{TATA}-lacZ* (available from Clontech, Palo Alto, CA). Y187 contains a lacZ Reporter Gene driven by GAL4 binding sites.

- 5 SFY526: *MAT α* , *ura3-52*, *his3-200*, *lys2-801*, *ade2-101*, *trp1-901*, *leu2-3,112*, *gal4-542*, *gal80-538*, *can'*, *URA3::GAL1-lacZ* (available from Clontech, Palo Alto, CA). SFY526 contains HIS3 and lacZ Reporter Genes driven by GAL4 binding sites.
- 10 HF7c: *MAT α* , *ura3-52*, *his3-200*, *lys2-801*, *ade2-101*, *trp1-901*, *leu2-3,112*, *gal4-542*, *gal80-538*, *LYS2::GAL1-HIS3*, *URA3::GAL1_{UAS17mers(x3)}-CYC1-lacZ* (available from Clontech, Palo Alto, CA). HF7c contains HIS3 and lacZ Reporter Genes driven by GAL4 binding sites.
- 15 YRG-2: *MAT α* , *ura3-52*, *his3-200*, *lys2-801*, *ade2-101*, *trp1-901*, *leu2-3,112*, *gal4-542*, *gal80-538*, *LYS2::GAL1_{UAS}-GAL1_{TATA}-HIS3*, *URA3::GAL1_{UAS17mers(x3)}-CYC1-lacZ* (available from Stratagene Cloning Systems, La Jolla, CA). YRG-2 contains HIS3 and lacZ Reporter Genes driven by GAL4 binding sites.
- 20 Many other strains commonly known and available in the art can be used.
 If not already lacking in endogenous Reporter Gene activity, cells mutant in the Reporter Gene may be selected by known methods, or the cells can be made mutant in the target Reporter Gene by known gene-disruption methods prior to introducing the Reporter Gene (Rothstein, 1983, Meth. Enzymol. 101:202-211).
- 25 In a specific embodiment, plasmids encoding the different fusion protein populations can be both introduced into a single host cell (e.g., a haploid yeast cell) containing one or more Reporter Genes, by co-transformation, to conduct the assay for protein-protein interactions. More preferably, the two fusion protein populations are introduced into a single cell either by mating (e.g., of yeast cells) or cell fusions (e.g., of
- 30 mammalian cells). In a mating type assay, conjugation of haploid yeast cells of opposite mating type that have been transformed, respectively, with a binding domain fusion expression construct (preferably a plasmid) and an activation (or inhibitor) domain fusion expression construct (preferably a plasmid), delivers both constructs into the same diploid cell. The mating type of a yeast strain may be manipulated by transformation with the HO
- 35 gene (Herskowitz and Jensen, 1991, Meth. Enzymol. 194:132-146).

In a specific embodiment, a yeast interaction mating assay is employed, using two different types of host cells, strain-types α and a , of the yeast *Saccharomyces cerevisiae*. The host cell preferably contains at least two Reporter Genes, each with one or more binding sites for the DNA-binding domain, *e.g.*, of a transcriptional activator. The
5 activator domain and DNA binding domain are each parts of chimeric proteins formed from the two respective populations of proteins. One set of host cells, for example the a strain cells, contains fusions of the library of nucleotide sequences with the DNA-binding domain of a transcriptional activator, such as GAL4. The hybrid proteins expressed in this set of host cells are capable of recognizing the DNA-binding site on the Reporter Gene. The
10 second set of yeast host cells, for example α strain cells, contains nucleotide sequences encoding fusions of a library of DNA sequences fused to the activation domain of a transcriptional activator.

In a specific embodiment, the fusion protein constructs are introduced into the host cell as a set of plasmids. These plasmids are preferably capable of autonomous
15 replication in a host yeast cell and preferably can also be propagated in *E. coli*. The plasmids contain a promoter directing the transcription of the DNA binding or activation domain fusion genes, and a transcriptional termination signal. The plasmids also preferably contain a selectable marker gene, permitting selection of cells containing the plasmids. The plasmids can be single-copy or multi-copy. Single-copy yeast plasmids that have the yeast
20 centromere may also be used to express the activation and DNA binding domain fusions (Elledge et al., 1988, Gene 70:303-312). In another embodiment, the fusion constructs are introduced directly into the yeast chromosome via homologous recombination. The homologous recombination for these purposes is mediated through yeast sequences that are not essential for vegetative growth of yeast, *e.g.*, the MER2, MER1, ZIPI, REC102, or
25 ME14 genes.

Bacteriophage vectors can also be used to express the DNA binding domain and/or activation domain fusion proteins. Libraries can generally be prepared faster and more easily from bacteriophage vectors than from plasmid vectors.

In a specific embodiment, the invention provides a method of detecting one
30 or more protein-protein interactions comprising (a) recombinantly expressing flt-4 or a derivative, homolog or analog thereof in a first population of yeast cells being of a first mating type and comprising a first fusion protein containing the flt-4 sequence and a DNA binding domain, wherein said first population of yeast cells contains a first nucleotide sequence operably linked to a promoter driven by one or more DNA binding sites
35 recognized by said DNA binding domain such that an interaction of said first fusion protein with a second fusion protein, said second fusion protein comprising a transcriptional

activation domain, results in increased transcription of said first nucleotide sequence; (b) negatively selecting to eliminate those yeast cells in said first population in which said increased transcription of said first nucleotide sequence occurs in the absence of said second fusion protein; (c) recombinantly expressing in a second population of yeast cells of a
5 second mating type different from said first mating type, a plurality of said second fusion proteins, each second fusion protein comprising a sequence of a fragment, derivative, homolog or analog of VEGF-C or VEGF-D and an activation domain of a transcriptional activator, in which the activation domain is the same in each said second fusion protein; (d) mating said first population of yeast cells with said second population of yeast cells to form
10 a third population of diploid yeast cells, wherein said third population of diploid yeast cells contains a second nucleotide sequence operably linked to a promoter driven by a DNA binding site recognized by said DNA binding domain such that an interaction of a first fusion protein with a second fusion protein results in increased transcription of said second nucleotide sequence, in which the first and second nucleotide sequences can be the same or
15 different; and (e) detecting said increased transcription of said first and/or second nucleotide sequence, thereby detecting an interaction between a first fusion protein and a second fusion protein.

In a specific embodiment, the bait flt-4 sequence and the prey library of chimeric genes are combined by mating the two yeast strains on solid media for a period of
20 approximately 6-8 hours. Optionally, the mating is performed in liquid media. The resulting diploids contain both kinds of chimeric genes, *i.e.*, the DNA-binding domain fusion and the activation domain fusion.

Preferred reporter genes include the URA3, HIS3 and/or the lacZ genes (*e.g.*, Rose and Botstein, 1983, Meth. Enzymol. 101:167-180) operably linked to GAL4 DNA-
25 binding domain recognition elements. Other reporter genes comprise the functional coding sequences for, but not limited to, Green Fluorescent Protein (GFP) (Cubitt et al., 1995, Trends Biochem. Sci. 20:448-455), luciferase, LEU2, LYS2, ADE2, TRP1, CAN1, CYH2, GUS, CUP1, or chloramphenicol acetyl transferase (CAT). Expression of LEU2, LYS2, ADE2 and TRP1 are detected by growth in a specific defined media; GUS and CAT can be
30 monitored by well known enzyme assays; and CAN1 and CYH2 are detected by selection in the presence of canavanine and cycloheximide, respectively. With respect to GFP, the natural fluorescence of the protein is detected.

In a specific embodiment, transcription of the Reporter Gene is detected by a linked replication assay. For example, as described by Vasavada et al., 1991, Proc. Natl.
35 Acad. Sci. USA 88:10686-10690, expression of SV40 large T antigen is under the control of the E1B promoter responsive to GAL4 binding sites. The replication of a plasmid

containing the SV40 origin of replication indicates the reconstruction of the GAL4 protein and a protein-protein interaction. Alternatively, a polyoma virus replicon can be employed (Vasavada et al., 1991, Proc. Natl. Acad. Sci. USA 88:10686-10690).

In another embodiment, the expression of Reporter Genes that encode
5 proteins can be detected by immunoassay, *i.e.*, by detecting the immunospecific binding of an antibody to such protein, which antibody can be labeled, or alternatively, which antibody can be incubated with a labeled binding partner to the antibody, so as to yield a detectable signal.

Alam and Cook, 1990, Anal. Biochem. 188:245-254 disclose non-limiting
10 examples of detectable marker genes that can be operably linked to a transcriptional regulatory region responsive to a reconstituted transcriptional activator, and thus can be used as Reporter Genes.

The activation of Reporter Genes like URA3 or HIS3 enables the cells to grow in the absence of uracil or histidine, respectively, and hence serves as selectable
15 markers. Thus, after mating, the cells exhibiting protein-protein interactions are selected for the ability to grow in media lacking a nutritional component, such as uracil or histidine, respectively (referred to as -URA (URA minus) and -HIS (HIS minus) medium, respectively). The -HIS medium preferably contains 3-amino-1,2,4-triazole (3-AT), which is a competitive inhibitor of the HIS3 gene product and thus requires higher levels of
20 transcription to achieve selection (Durfee et al., 1993, Genes Dev. 7:555-569). Similarly, 6-azauracil, which is an inhibitor of the URA3 gene product, can be included in -URA medium (Le Douarin et al., 1995, Nucl. Acids Res. 23:876-878). URA3 gene activity can also be detected and/or measured by determining the activity of its gene product, orotidine-5'-monophosphate decarboxylase (Pierrat et al., 1992, Gene 119:237-245; Wolcott et al.,
25 1966, Biochem. Biophys. Acta 122:532-534). In other embodiments of the invention, the activities of the reporter genes like lacZ or GFP are monitored by measuring a detectable signal (*e.g.*, chromogenic or fluorescent, respectively) that results from the activation of these Reporter Genes. For example, lacZ transcription can be monitored by incubation in the presence of a chromogenic substrate, such as X-gal (5-bromo-4-chloro-3-indolyl- β -D-galactoside), for its encoded enzyme, β -galactosidase. The pool of all interacting proteins isolated in this manner by mating the *flt-4* sequence product and the library identifies the
30 "*flt-4* interactive population".

In a specific embodiment of the invention, false positives arising from transcriptional activation by the DNA binding domain fusion proteins in the absence of a
35 transcriptional activator domain fusion protein are prevented or reduced by negative selection for such activation within a host cell containing the DNA binding fusion

population, prior to exposure to the activation domain fusion population. By way of example, if such cell contains URA3 as a Reporter Gene, negative selection is carried out by incubating the cell in the presence of 5-fluoroorotic acid (5-FOA) which kills URA+ cells (Rothstein, 1983, Meth. Enzymol. 101:167-180). Hence, if the DNA-binding domain fusions by themselves activate transcription, the metabolism of 5-FOA will lead to cell death and the removal of self-activating DNA-binding domain hybrids.

Negative selection involving the use of a selectable marker as a Reporter Gene and the presence in the cell medium of an agent toxic or growth inhibitory to the host cells in the absence of Reporter Gene transcription is preferred, since it allows a higher rate of processing than other methods. As will be apparent, negative selection can also be carried out on the activation domain fusion population prior to interaction with the DNA binding domain fusion population, by similar methods, either alone or in addition to negative selection of the DNA binding fusion population.

Negative selection can also be carried out on the recovered flt-4:VEGF-C/D pairs by known methods (*e.g.*, Bartel et al., 1993, BioTechniques 14:920-924) although pre-negative selection (prior to the interaction assay), as described above, is preferred. For example, each plasmid encoding a protein or peptide or polypeptide fused to the activation domain (one-half of a detected interacting pair) can be transformed back into the original screening strain, either alone or with a plasmid encoding only the DNA-binding domain, the DNA-binding domain fused to the detected interacting protein, or the DNA-binding domain fused to a protein that does not affect transcription or participate in the protein-protein interaction; a positive interaction detected with any plasmid other than that encoding the DNA-binding domain fusion to the detected interacting protein indicates that the activation domain yields false positives, and it is subsequently eliminated from the screen.

In a specific embodiment, the flt-4 plasmid population is transformed in a yeast strain of a first mating type (α or alpha), and the second plasmid population (containing the library of DNA sequences) is transformed in a yeast strain of different mating type. Both strains are preferably mutant for URA3 and HIS3, and contain URA3, and optionally lacZ, as a Reporter Genes. The first set of yeast cells are positively selected for the flt-4 plasmids and are negatively selected for false positives by incubation in medium lacking the selectable marker (*e.g.*, uracil) and containing 5-FOA. Yeast cells of the second mating type are transformed with the second plasmid population, and are positively selected for the presence of the plasmids containing the library of fusion proteins. Selected cells are pooled. Both groups of pooled cells are mixed together and mating is allowed to occur on a solid phase. The resulting diploid cells are then transferred to

selective media that selects for the presence of each plasmid and for activation of Reporter Genes.

In a specific embodiment of the invention, after an interactive population is obtained, the DNA sequences encoding the pairs of interactive proteins are isolated by a method wherein either the DNA-binding domain hybrids or the activation domain hybrids are amplified, in separate respective reactions. Preferably, the amplification is carried out by polymerase chain reaction (PCR) (U.S. Patent Nos. 4,683,202, 4,683,195 and 4,889,818; Gyllenstein et al., 1988, Proc. Natl. Acad. Sci. USA 85:7652-7656; Ochman et al., 1988, Genetics 120:621-623; Loh et al., 1989, Science 243:217-220; Innis et al., 1990, PCR Protocols, Academic Press, Inc., San Diego, CA), using pairs of oligonucleotide primers specific for either the DNA-binding domain hybrids or the activation domain hybrids. This PCR reaction can also be performed on pooled cells expressing interacting protein pairs, preferably as pooled arrays of interactants. Other amplification methods known in the art can be used, including but not limited to ligase chain reaction (EP 320,308), use of Q β replicase, or methods listed in Kricka et al., 1995, Molecular Probing, Blotting, and Sequencing, Chap. 1 and Table IX, Academic Press, New York.

The plasmids encoding the DNA-binding domain hybrid proteins and the activation domain hybrid proteins can also be isolated and cloned by any of the methods well known in the art. For example, but not by way of limitation, if a shuttle (yeast to *E. coli*) vector is used to express the fusion proteins, the genes can be recovered by transforming the yeast DNA into *E. coli* and recovering the plasmids from *E. coli* (e.g., Hoffman et al., 1987, Gene 57:267-272). Alternatively, the yeast vector can be isolated, and the insert encoding the fusion protein subcloned into a bacterial expression vector, for growth of the plasmid in *E. coli*.

5.5. PHARMACEUTICAL COMPOSITIONS AND THERAPEUTIC/PROPHYLACTIC ADMINISTRATION

The invention provides methods of treatment (and prophylaxis) by administration to a subject of an effective amount of a Therapeutic of the invention. In a preferred aspect, the Therapeutic is substantially purified. The subject is preferably an animal including, but not limited to animals such as cows, pigs, horses, chickens, cats, dogs, etc., and is preferably a mammal, and most preferably human. In a specific embodiment, a non-human mammal is the subject.

Formulations and methods of administration that can be employed when the Therapeutic comprises a nucleic acid are described in Sections 5.2.1 and 5.2.2, *supra*; additional appropriate formulations and routes of administration can be selected from among those described herein below.

Various delivery systems are known and can be used to administer a Therapeutic of the invention, *e.g.*, encapsulation in liposomes, microparticles, and microcapsules; use of recombinant cells capable of expressing the Therapeutic, use of receptor-mediated endocytosis (*e.g.*, Wu and Wu, 1987, *J. Biol. Chem.* 262:4429-4432);
5 construction of a Therapeutic nucleic acid as part of a retroviral or other vector, etc. Methods of introduction include but are not limited to intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, epidural, and oral routes. The compounds may be administered by any convenient route, for example by infusion, by bolus injection, by absorption through epithelial or mucocutaneous linings (*e.g.*, oral, rectal
10 and intestinal mucosa, etc.), and may be administered together with other biologically active agents. Administration can be systemic or local. In addition, it may be desirable to introduce the pharmaceutical compositions of the invention into the central nervous system by any suitable route, including intraventricular and intrathecal injection; intraventricular injection may be facilitated by an intraventricular catheter, for example, attached to a
15 reservoir, such as an Ommaya reservoir. Pulmonary administration can also be employed, *e.g.*, by use of an inhaler or nebulizer, and formulation with an aerosolizing agent.

In a specific embodiment, it may be desirable to administer the pharmaceutical compositions of the invention locally to the area in need of treatment. This may be achieved by, for example, and not by way of limitation, local infusion during
20 surgery, topical application, *e.g.*, in conjunction with a wound dressing after surgery, by injection, by means of a catheter, by means of a suppository, or by means of an implant, said implant being of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, or fibers. In one embodiment, administration can be by direct injection at the site (or former site) of a malignant tumor or neoplastic or pre-neoplastic
25 tissue.

In another embodiment, the Therapeutic can be delivered in a vesicle, in particular a liposome (Langer, 1990, *Science* 249:1527-1533; Treat et al., 1989, In: *Liposomes in the Therapy of Infectious Disease and Cancer*, Lopez-Berestein and Fidler, eds., Liss, New York, pp. 353-365; Lopez-Berestein, *ibid.*, pp. 317-327; see generally *ibid.*)
30 In yet another embodiment, the Therapeutic can be delivered via a controlled release system. In one embodiment, a pump may be used (Langer, *supra*; Sefton, 1987, *CRC Crit. Ref. Biomed. Eng.* 14:201-240; Buchwald et al., 1980, *Surgery* 88:507-516; Saudek et al., 1989, *N. Engl. J. Med.* 321:574-579). In another embodiment, polymeric materials can be used (*Medical Applications of Controlled Release*, Langer and Wise, eds.,
35 CRC Press, Boca Raton, Florida, 1974; *Controlled Drug Bioavailability, Drug Product Design and Performance*, Smolen and Ball, eds., Wiley, New York, 1984; Ranger and

Peppas, 1983, *Macromol. Sci. Rev. Macromol. Chem.* 23:61; Levy et al., 1985, *Science* 228:190-192; During et al., 1989, *Ann. Neurol.* 25:351-356; Howard et al., 1989, *J. Neurosurg.* 71:858-863). In yet another embodiment, a controlled release system can be placed in proximity of the therapeutic target, *i.e.*, the brain, thus requiring only a fraction of the systemic dose (*e.g.*, Goodson, 1984, In: *Medical Applications of Controlled Release*, supra, Vol. 2, pp. 115-138). Other controlled release systems are discussed in the review by Langer (1990, *Science* 249:1527-1533).

In a specific embodiment where the Therapeutic is a nucleic acid encoding a protein Therapeutic, the nucleic acid can be administered *in vivo* to promote expression of its encoded protein, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, *e.g.*, by use of a retroviral vector (U.S. Patent No. 4,980,286), or by direct injection, or by use of microparticle bombardment (*e.g.*, a gene gun; Biolistic, Dupont), or by coating it with lipids, cell-surface receptors or transfecting agents, or by administering it in linkage to a homeobox-like peptide which is known to enter the nucleus (*e.g.*, Joliot et al., 1991, *Proc. Natl. Acad. Sci. USA* 88:1864-1868), etc. Alternatively, a nucleic acid Therapeutic can be introduced intracellularly and incorporated by homologous recombination within host cell DNA for expression.

The present invention also provides pharmaceutical compositions. Such compositions comprise a therapeutically effective amount of a Therapeutic, and a pharmaceutically acceptable carrier. In a specific embodiment, the term "pharmaceutically acceptable" means approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly, in humans. The term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the therapeutic is administered. Such pharmaceutical carriers can be sterile liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, including but not limited to peanut oil, soybean oil, mineral oil, sesame oil and the like. Water is a preferred carrier when the pharmaceutical composition is administered orally. Saline and aqueous dextrose are preferred carriers when the pharmaceutical composition is administered intravenously. Saline solutions and aqueous dextrose and glycerol solutions are preferably employed as liquid carriers for injectable solutions. Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like. The composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions,

suspensions, emulsions, tablets, pills, capsules, powders, sustained-release formulations and the like. The composition can be formulated as a suppository, with traditional binders and carriers such as triglycerides. Oral formulation can include standard carriers such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, etc. Examples of suitable pharmaceutical carriers are described in "Remington's Pharmaceutical Sciences" by E.W. Martin. Such compositions will contain a therapeutically effective amount of the Therapeutic, preferably in purified form, together with a suitable amount of carrier so as to provide the form for proper administration to the patient. The formulation should suit the mode of administration.

In a preferred embodiment, the composition is formulated, in accordance with routine procedures, as a pharmaceutical composition adapted for intravenous administration to human beings. Typically, compositions for intravenous administration are solutions in sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lignocaine to ease pain at the site of the injection. Generally, the ingredients are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water-free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent. Where the composition is to be administered by infusion, it can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the composition is administered by injection, an ampoule of sterile water or saline for injection can be provided so that the ingredients may be mixed prior to administration.

The Therapeutics of the invention can be formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with free carboxyl groups such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., those formed with free amine groups such as those derived from isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc., and those derived from sodium, potassium, ammonium, calcium, and ferric hydroxides, etc.

The amount of the Therapeutic of the invention which will be effective in the treatment of a particular disorder or condition will depend on the nature of the disorder or condition, and can be determined by standard clinical techniques. In addition, *in vitro* assays may optionally be employed to help identify optimal dosage ranges. The precise dose to be employed in the formulation will also depend on the route of administration, and the seriousness of the disease or disorder, and should be decided according to the judgment of the practitioner and each patient's circumstances. However, suitable dosage ranges for intravenous administration are generally about 20-500 micrograms of active compound per kilogram body weight. Suitable dosage ranges for intranasal administration are generally

about 0.01 pg/kg body weight to 1 mg/kg body weight. Effective doses may be extrapolated from dose-response curves derived from *in vitro* or animal model test systems.

Suppositories generally contain active ingredient in the range of 0.5% to 10% by weight; oral formulations preferably contain 10% to 95% active ingredient.

5 The invention also provides a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients of the pharmaceutical compositions of the invention. Optionally associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, which notice reflects approval by the agency of
10 manufacture, use or sale for human administration.

5.6. ANIMAL MODELS

The present invention also provides animal models. In one embodiment, animal models for diseases and disorders involving flt-4:VEGF-C/D complexes are
15 provided. These include, but are not limited to, secondary prostate tumor metastases. Such animals can be initially produced by promoting homologous recombination or insertional mutagenesis between flt-4 and VEGF-C or VEGF-D genes in the chromosome, and exogenous flt-4 and VEGF-C or VEGF-D genes that have been rendered biologically inactive or deleted (preferably by insertion of a heterologous sequence, *e.g.*, an antibiotic
20 resistance gene). In a preferred aspect, homologous recombination is carried out by transforming embryo-derived stem (ES) cells with a vector containing the insertionally inactivated flt-4 and VEGF-C or VEGF-D gene, such that homologous recombination occurs, followed by injecting the transformed ES cells into a blastocyst, and implanting the blastocyst into a foster mother, followed by the birth of the chimeric animal ("knockout
25 animal") in which a flt-4 and/or VEGF-C or VEGF-D gene has been inactivated or deleted (Capecchi, 1989, Science 244:1288-1292). The chimeric animal can be bred to produce additional knockout animals. Such animals can be mice, hamsters, sheep, pigs, cattle, etc., and are preferably non-human mammals. In a specific embodiment, a knockout mouse is produced.

30 In a different embodiment of the invention, transgenic animals that have incorporated and express (or overexpress or mis-express) a functional flt-4 and/or VEGF-C or VEGF-D gene, *e.g.* by introducing the flt-4 and VEGF-C or VEGF-D genes under the control of a heterologous promoter (*i.e.*, a promoter that is not the native flt-4 or VEGF-C or VEGF-D promoter) that either overexpresses the protein or proteins, or expresses them in
35 tissues not normally expressing the complexes or proteins, can have use as animal models of diseases and disorders characterized by elevated levels of flt-4:VEGF-C/D complexes.

Such animals can be used to screen or test molecules for the ability to treat or prevent the diseases and disorders cited *supra*.

In one embodiment, the invention provides a recombinant non-human animal containing both a *flt-4* gene and a VEGF-C or VEGF-D gene in which the *flt-4* gene is under the control of a promoter that is not the native *flt-4* gene promoter and the VEGF-C or VEGF-D gene is under the control of a promoter that is not the native VEGF-C or VEGF-D gene promoter.

The following series of examples are presented by way of illustration and not by way of limitation on the scope of the present invention.

6. EXAMPLES

The following experiments demonstrate that *flt-4* is expressed only in prostate cancer cells that have metastatic potential or are or are derived from secondary prostate tumor metastases of a primary prostate tumor, such that *flt-4* expression in a prostate cancer cell can serve as a diagnostic and prognostic marker of prostate cancer, and further that methods inhibiting expression or activity of *flt-4* are useful in the treatment, inhibition or prevention of prostate cancer.

6.1. FLT-4 EXPRESSION DETERMINED BY IMMUNOHISTOCHEMISTRY

Representative tissue samples of benign prostate hyperplasia, primary prostate carcinoma, normal lymph nodes and secondary prostate tumor lymphatic metastasis were stained with an anti-*flt-4* antibody (Santa Cruz Corp., Santa Cruz, CA) using the Zemed Histostain kit obtained from Zemed (So. San Francisco, CA). The immunohistochemical assay was carried out by sequential application of the diluted primary antibody for 30 minutes, biotinylated secondary antibody for 15 minutes and HRP-Streptavidin for 15 minutes. Immunoreactivity was visualized with either AEC (3-amino-9-ethylcarbazole) or DAB (3,3'-diaminobenzidine) chromogens and sections were counterstained with hematoxylin. Appropriate negative controls were performed using rabbit or goat or isotyped matched mouse antibodies and all negative controls showed very low background. Positive and negative controls were run in parallel with each batch. The results are presented in Figs. 2A-2F.

Figs. 2A and 2B, normal lymph node tissue and benign prostate hyperplasia tissue, respectively, no *flt-4* expression is seen. However, *flt-4* expression is evident in lymph node tissue with prostatic metastases (Figs. 2C and 2D). Fig. 2E clearly shows the

difference in flt-4 expression between prostate cancer tissue and benign prostate hyperplasia tissue. Fig. 2F also clearly demonstrates that flt-4 is expressed in prostate cancer tissue.

In the clinical samples tested, all benign epithelium tested (5/5) negative for flt-4 expression. All benign lymph nodes (7/7) also tested negative for flt-4 expression. Most prostate carcinoma (9/10) also tested negative for flt-4 expression. Interestingly, the only exception for prostate carcinoma was obtained from a patient who developed local recurring disease subsequent to radical prostatectomy. All prostate cancer metastases in lymph nodes tested (2/2) positive for flt-4 expression. These same samples all tested positive for VEGF-C expression, indicating that the secondary prostate tumor metastases exhibit a VEGF-C/flt-4 autocrine loop.

Human prostate cancer cell lines were also tested for flt-4, VEGF-C, flk-1 and VEGF expression by immunocytochemistry. The human prostate cancer cell lines, LNCaP, PC-3, DU145, and TSUPr1, were grown in a slide chamber and were fixed with 10% formalin and stained with anti-flt-4 antibody as described above. The antibodies to VEGF-C, VEGF and flk-1 were also obtained from Santa Cruz Corp. (Santa Cruz, CA). The results, that flt-4 is expressed in each human prostate cancer cell line, are shown in Figs. 3A-3D.

6.2. FLT-4 EXPRESSION DETERMINED BY REVERSE TRANSCRIPTASE PCR

Total RNA was isolated from human prostate cancer cell lines LNCaP, a hormone sensitive cell line, PC-3, DU-145 and TSUPr1, hormone independent cell lines, using a RNazol B kit (Biotex, Houston, TX). After RNase-free DNase I treatment (Promega, Madison, WI), 5 µg of RNA was reverse transcribed using Superscript II (Gibco-BRL, London, UK).

Polymerase chain reaction ("PCR") was performed on the reverse transcribed cDNA samples using primers for VEGF and its receptor flk-1 and VEGF-C and its receptor flt-4. The primers used are as follows:

VEGF: 5'-CGAAGTGGTGAAGTTCATGGATG-3' (SEQ ID NO:4) and 5'-TTCTGTATCAGTCTTTCCTGGTGAG-3' (SEQ ID NO:5),

flk-1: 5'-CTGGCATGGTCTTCTGTGAAAGCA-3' (SEQ ID NO:6) and 5'-AATACCAGTGGATGTGATGCGG-3' (SEQ ID NO:7),

VEGF-C: 5'-TACCACAGTGTGAGGCAGCG-3' (SEQ ID NO:8) and 5'-ATCAAATTCTCGGTTGGCCC-3', (SEQ ID NO:9),

flt-4: 5'-AGAGGGATGGAGTTCCTGGC-3' (SEQ ID NO:10) and 5'-AATACCAGTGGATGTGATGCGG-3' (SEQ ID NO:11).

PCR was performed using Biolase *Taq* polymerase (Bioline, London, UK) with the supplied buffer and 3 mM MgCl₂ for VEGF, VEGF-C and flt-4 or with the supplied buffer and 3.5 mM MgCl₂ for flk-1. The amplification conditions were 35 cycles of denaturation at 94°C for 30 seconds; annealing at 63°C for VEGF, 62°C for flk-1, 57°C for VEGF-C and 63°C for flt-4 for 1 minute; and extension at 72°C for 1 minute. After the 35 cycles, a final extension step at 72°C for 10 minutes is performed. The amplification products were separated on 2.5% agarose gels. The results are shown in Fig. 4.

As shown in Fig. 4, flt-4 is expressed in all the human prostate cancer cell lines tested. In addition, the ligand for flt-4, VEGF-C, is also expressed in all the hormone independent human prostate cancer cell lines tested. These results agree with the immunohistochemistry results and is consistent with the presence of a VEGF-C/flt-4 autocrine loop in metastatic prostate cancer cells.

6.3. FLOW CYTOMETRY

LNCaP cells (1×10^6) were pelleted and incubated with rabbit anti-flt-4 serum or normal rabbit serum (negative control), each obtained from Santa Cruz Corp. (Santa Cruz, CA), for 30 minutes at 4°C. Cells were washed once with PBS and then incubated with biotinylated anti-rabbit antibody for 30 minutes at 4°C. After another wash with PBS, cells were incubated with strep-PE for 30 minutes at 4°C. Cells were washed once more with PBS and analyzed on a Becton-Dickinson FACSCalibur flow cytometer. The LNCaP cell line was derived from a secondary prostate metastasis in lymph node tissue. The results are shown in Fig. 5.

Fig. 5 clearly shows that flt-4 expression can be detected on the surface of live LNCaP cells.

6.4. INHIBITION OF PC-3 GROWTH USING ANTISENSE OLIGONUCLEOTIDES

This example demonstrates that antisense oligonucleotides for inhibiting the expression of VEGF and its receptors flk-1 and flt-1 and VEGF-C and its receptor flt-4 inhibited the growth of the hormone independent human prostate cancer cell line PC-3, a cell line derived from a prostate tumor metastasis.

PC-3 cells were seeded at 2000 cells/well in a 96 well plate or seeded at 20,000 cells/well in a 24 well plate in serum free X-VIVO medium (Biowhittaker, Walkerville, MD). Antisense oligos were added to the cells at increasing concentration of 1 to 100 μ M. Media and oligos were refreshed every third day and the cultures were maintained for 14 days. Viable cells were quantitated either by hemacytometer cell-

counting or by a MTS 96-well viable cell assay obtained from Promega (Madison, WI). Cell proliferation using the MTS assay was expressed as a percentage of control. The experiments were done in duplicate. The sequences of the oligonucleotides were as follows:

- 5 Anti-VEGF: 5'-AGACAGCAGAAAGTTCATGGT-3' (SEQ ID NO:12)
 Anti-VEGF-C: 5'-CAAGTGCATGGTGGA-3' (SEQ ID NO:13)
 Anti-flk-1: 5'-CACCTTGCTCTGCAT-3' (SEQ ID NO:14)
 Anti-flt-1: 5'-CCCGGTGTCCCAGA-3' (SEQ ID NO:15)
 Anti-flt-4: 5'-GGCGCCCCGCTGCAT-3' (SEQ ID NO:3)
10 Control oligo: 5'-TACGTAGTATGGTGTAC-3' (SEQ ID NO:16)

The results are shown in Fig. 6.

An oligo concentration of 100 μ M proved to be toxic to the cells and is excluded from Fig. 6. As indicated in Fig. 6, growth inhibition was generally maximal at 5-10 μ M. Repression of flt-1 and flt-4 expression had the greatest inhibitory effect on PC-3 cell
15 growth, approximately 50%, followed by VEGF-C, approximately 20-30%. Anti-VEGF and anti-flk-1 oligos inhibited growth by approximately 20% as compared to control oligo. The differential inhibition between VEGF-C/flt-4 and VEGF/flk-1 may reflect the relative importance of each autocrine pathway. Thus, these results demonstrate that anti-flt-4 and anti-VEGF-C antisense oligonucleotides are effective for inhibiting or suppressing prostate
20 cancer metastases.

The invention claimed and described herein is not to be limited in scope by the specific embodiments herein disclosed since these embodiments are intended as illustrations of several aspects of the invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those
25 skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims.

A number of references are cited herein, the entire disclosures of which are incorporated herein, in their entirety, by reference.

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WE CLAIM:

1. A method for detection of metastatic potential comprising detecting expression of flt-4 in a prostate cell, wherein expression of flt-4 indicates that said cell has metastatic
5 potential.
2. A method for detection of metastatic potential comprising identifying a prostate cell in a body fluid sample obtained from a subject and detecting expression of flt-4 in said cell, wherein expression of flt-4 indicates that said cell is a prostate cancer cell that has
10 metastatic potential or is a secondary prostate tumor metastasis, or is derived therefrom.
3. The method according to claim 1 or 2 in which the prostate cell is identified by using an antibody or a portion thereof that binds to a prostate cell-specific marker.
- 15 4. The method according to claim 3 in which the prostate cell-specific marker is selected from the group consisting of prostate-specific antigen (PSA), prostate-specific membrane antigen (PSMA), prostate secretory protein (PSP), prostate acid phosphatase (PAP), human glandular kallekrein 2 (HK-2), prostate stem cell antigen (PSCA) and PTI-1.
- 20 5. The method according to claim 1 or 2 in which flt-4 expression is detected using an antibody or a portion thereof that binds to flt-4.
6. The method according to claim 1 or 2 in which flt-4 expression is detected using a nucleic acid molecule, said molecule comprising a nucleotide sequence consisting of a
25 sequence of at least 6 contiguous nucleotides complementary to the nucleotide sequence set forth in SEQ ID NO:1.
7. The method according to claim 2 in which the body fluid is blood, urine or semen.
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8. The method according to claim 2 in which identifying the prostate cell and detecting flt-4 expression are performed simultaneously.
9. The method according to claim 8 in which any immunofluorescence assay is
35 employed.

10. The method according to claim 9 in which the immunofluorescence assay employs a flow cytometer or a laser scanning cytometer.

5 11. A method for diagnosing metastatic prostate cancer in a subject comprising identifying a prostate cell in a body fluid sample obtained from the subject and detecting expression of flt-4 in the prostate cell, wherein expression of flt-4 in a prostate cell indicates that the subject has metastatic prostate cancer.

10 12. A method for determining the prognosis of a subject with prostate cancer comprising identifying a prostate cell in a body fluid sample obtained from a subject with prostate cancer and detecting expression of flt-4 in the prostate cell, wherein expression of flt-4 in said cell indicates that the subject has a worse prognosis as compared to a second subject in whose prostate cell no flt-4 expression or activity is detected.

15 13. A method of treating, inhibiting or preventing a secondary prostate tumor metastasis comprising administering to a subject in which such treatment, inhibition or prevention is desired a therapeutically effective amount of a molecule that inhibits flt-4 expression or activity.

20 14. The method according to claim 13 in which the molecule is a protein comprising a fragment of flt-4, which fragment consists of at least the amino acid sequence set forth in SEQ ID NO:2, which protein acts as a competitive inhibitor of flt-4 binding to its ligand VEGF-C.

25 15. The method according to claim 14 in which the protein is soluble.

16. The method according to claim 13 in which the molecule is a nucleic acid molecule comprising a nucleotide sequence which encodes for a fragment of flt-4, which fragment acts as a competitive inhibitor of flt-4 binding to its ligand VEGF-C.

30 17. The method according to claim 13 in which the molecule is an antisense oligonucleotide comprising a nucleotide sequence consisting of at least 6 contiguous nucleotides complementary to the nucleotide sequence set forth in SEQ ID NO:1.

35 18. The method according to claim 17 in which the antisense oligonucleotide comprises the nucleotide sequence 5'-GGCGCCCCGCTGCAT-3' (SEQ ID NO:3).

19. The method according to claim 13 in which the molecule is an antibody or a portion thereof that binds to flt-4.

20. A method for screening for a molecule that treats, inhibits or prevents a secondary prostate tumor metastasis comprising contacting a prostate cell that expresses flt-4 with a candidate molecule and comparing the level of flt-4 expression in the cell so contacted with a prostate cell expressing flt-4 not so contacted, wherein a lower level of flt-4 expression in the contacted cell as compared to the non-contacted cell indicates that the candidate molecule has activity in treating, inhibiting or preventing secondary prostate tumor metastases.

21. A method for screening for a molecule that treats, inhibits or prevents a secondary prostate tumor metastasis comprising measuring the levels of complex formed from flt-4 and VEGF-C in the presence of a candidate molecule under conditions conducive to the formation of said complex; and comparing levels of said complex that are formed in the absence of the molecule, wherein a lower level of said complex in the presence of the molecule indicates that the candidate molecule has activity in treating, inhibiting or preventing secondary prostate tumor metastases.

22. A method for screening for a molecule that treats, inhibits or prevents a secondary prostate tumor metastasis comprising measuring the levels of complex formed from flt-4 and VEGF-D in the presence of a candidate molecule under conditions conducive to the formation of said complex; and comparing levels of said complex that are formed in the absence of the molecule, wherein a lower level of said complex in the presence of the molecule indicates that the candidate molecule has activity in treating, inhibiting or preventing secondary prostate tumor metastases.

23. A method of monitoring the efficacy of a method of treatment or inhibition of metastatic prostate cancer comprising measuring the level of expression or activity of flt-4 in prostate cells obtained from a subject wherein said sample is taken from said subject after the application of said method and compared to (a) said level in a sample taken from said subject prior to the application of said method or (b) a standard level associated with the pretreatment stage of metastatic prostate cancer, in which a decrease in the level of flt-4 expression or activity in said sample taken after application of said method relative to the level of flt-4 expression or activity in said sample taken before application of said method or to said standard level indicates that said method is effective.

24. A pharmaceutical composition comprising a protein, which protein comprises a fragment of flt-4, which fragment consists of at least the amino acid sequence set forth in SEQ ID NO:2, which protein acts as a competitive inhibitor of flt-4 binding to its ligand VEGF-C; and a pharmaceutically acceptable carrier.

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25. The composition according to claim 24 in which the protein is soluble.

26. A pharmaceutical composition comprising a nucleic acid, which nucleic acid comprises a nucleotide sequence which encodes for a fragment of flt-4, which fragment acts
10 as a competitive inhibitor of flt-4 binding to its ligand VEGF-C; and a pharmaceutically acceptable carrier.

27. A pharmaceutical composition comprising an antisense oligonucleotide, which oligonucleotide comprises a nucleotide sequence consisting of at least 6 contiguous
15 nucleotides complementary to the nucleotide sequence set forth in SEQ ID NO:1; and a pharmaceutically acceptable carrier.

28. The composition according to claim 27 in which the antisense oligonucleotide comprises the nucleotide sequence 5'-GGCGCCCCGCTGCAT-3' (SEQ ID NO:3).

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29. A pharmaceutical composition comprising an antibody or a portion thereof that binds to flt-4; and a pharmaceutically acceptable carrier.

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ACCCACGCGC	AGCGGCCGGA	G	ATG	CAG	CGG	GGC	GCC	GCG	CTG	TGC	CTG	CGA	51			
			Met	Gln	Arg	Gly	Ala	Ala	Leu	Cys	Leu	Arg				
			1				5					10				
CTG	ACC	CCC	CCG	ACC	TTG	AAC	ATC	ACG	GAG	GAG	TCA	CAC	GTC	ATC	GAC	99
Leu	Trp	Leu	Cys	Leu	Gly	Leu	Leu	Asp	Gly	Leu	Val	Ser	Asp	Tyr	Ser	
				15					20					25		
ATG	ACC	CCC	CCG	ACC	TTG	AAC	ATC	ACG	GAG	GAG	TCA	CAC	GTC	ATC	GAC	147
Met	Thr	Pro	Pro	Thr	Leu	Asn	Ile	Thr	Glu	Glu	Ser	His	Val	Ile	Asp	
								35					40			
ACC	GGT	GAC	AGC	CTG	TCC	ATC	TCC	TGC	AGG	GGA	CAG	CAC	CCC	CTC	GAG	195
Thr	Gly	Asp	Ser	Leu	Ser	Ile	Ser	Cys	Arg	Gly	Gln	His	Pro	Leu	Glu	
		45					50					55				
TGG	GCT	TGG	CCA	GGA	GCT	CAG	GAG	GCG	CCA	GCC	ACC	GGA	GAC	AAG	GAC	243
Trp	Ala	Trp	Pro	Gly	Ala	Gln	Glu	Ala	Pro	Ala	Thr	Gly	Asp	Lys	Asp	
	60					65					70					
AGC	GAG	GAC	ACG	GGG	GTG	GTG	CGA	GAC	TGC	GAG	GGC	ACA	GAC	GCC	AGG	291
Ser	Glu	Asp	Thr	Gly	Val	Val	Arg	Asp	Cys	Glu	Gly	Thr	Asp	Ala	Arg	
75					80					85					90	
CCC	TAC	TGC	AAG	GTG	TTG	CTG	CTG	CAC	GAG	GTA	CAT	GCC	AAC	GAC	ACA	339
Pro	Tyr	Cys	Lys	Val	Leu	Leu	Leu	His	Glu	Val	His	Ala	Asn	Asp	Thr	
				95					100					105		
GGC	AGC	TAC	GTC	TGC	TAC	TAC	AAG	TAC	ATC	AAG	GCA	CGC	ATC	GAG	GGC	387
Gly	Ser	Tyr	Val	Cys	Tyr	Tyr	Lys	Tyr	Ile	Lys	Ala	Arg	Ile	Glu	Gly	
			110					115					120			
ACC	ACG	GCC	GCC	AGC	TCC	TAC	GTG	TTC	GTG	AGA	GAC	TTT	GAG	CAG	CCA	435
Thr	Thr	Ala	Ala	Ser	Ser	Tyr	Val	Phe	Val	Arg	Asp	Phe	Glu	Gln	Pro	
		125					130					135				
TTC	ATC	AAC	AAG	CCT	GAC	ACG	CTC	TTG	GTC	AAC	AGG	AAG	GAC	GCC	ATG	483
Phe	Ile	Asn	Lys	Pro	Asp	Thr	Leu	Leu	Val	Asn	Arg	Lys	Asp	Ala	Met	
	140					145					150					
TGG	GTG	CCC	TGT	CTG	GTG	TCC	ATC	CCC	GGC	CTC	AAT	GTC	ACG	CTG	CGC	531
Trp	Val	Pro	Cys	Leu	Val	Ser	Ile	Pro	Gly	Leu	Asn	Val	Thr	Leu	Arg	
155					160					165					170	
TCG	CAA	AGC	TCG	GTG	CTG	TGG	CCA	GAC	GGG	CAG	GAG	GTG	GTG	TGG	GAT	579
Ser	Gln	Ser	Ser	Val	Leu	Trp	Pro	Asp	Gly	Gln	Glu	Val	Val	Trp	Asp	
				175					180					185		
GAC	CGG	CGG	GGC	ATG	CTC	GTG	TCC	ACG	CCA	CTG	CTG	CAC	GAT	GCC	CTG	627
Asp	Arg	Arg	Gly	Met	Leu	Val	Ser	Thr	Pro	Leu	Leu	His	Asp	Ala	Leu	
			190					195					200			
TAC	CTG	CAG	TGC	GAG	ACC	ACC	TGG	GGA	GAC	CAG	GAC	TTC	CTT	TCC	AAC	675
Tyr	Leu	Gln	Cys	Glu	Thr	Thr	Trp	Gly	Asp	Gln	Asp	Phe	Leu	Ser	Asn	
		205					210					215				

FIG. 1A

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CCC Pro	TTC Phe 220	CTG Leu	GTG Val	CAC His	ATC Ile	ACA Thr 225	GGC Gly	AAC Asn	GAG Glu	CTC Leu	TAT Tyr 230	GAC Asp	ATC Ile	CAG Gln	CTG Leu	723
TTG Leu 235	CCC Pro	AGG Arg	AAG Lys	TCG Ser	CTG Leu 240	GAG Glu	CTG Leu	CTG Leu	GTA Val	GGG Gly 245	GAG Glu	AAG Lys	CTG Leu	GTC Val	CTC Leu 250	771
AAC Asn	TGC Cys	ACC Thr	GTG Val	TGG Trp 255	GCT Ala	GAG Glu	TTT Phe	AAC Asn	TCA Ser 260	GGT Gly	GTC Val	ACC Thr	TTT Phe	GAC Asp 265	TGG Trp	819
GAC Asp	TAC Tyr	CCA Pro	GGG Gly 270	AAG Lys	CAG Gln	GCA Ala	GAG Glu	CGG Arg 275	GGT Gly	AAG Lys	TGG Trp	GTG Val	CCC Pro 280	GAG Glu	CGA Arg	867
CGC Arg	TCC Ser	CAA Gln 285	CAG Gln	ACC Thr	CAC His	ACA Thr	GAA Glu 290	CTC Leu	TCC Ser	AGC Ser	ATC Ile	CTG Leu 295	ACC Thr	ATC Ile	CAC His	915
AAC Asn	GTC Val 300	AGC Ser	CAG Gln	CAC His	GAC Asp	CTG Leu 305	GGC Gly	TCG Ser	TAT Tyr	GTG Val	TGC Cys 310	AAG Lys	GCC Ala	AAC Asn	AAC Asn	963
GGC Gly 315	ATC Ile	CAG Gln	CGA Arg	TTT Phe	CGG Arg 320	GAG Glu	AGC Ser	ACC Thr	GAG Glu	GTC Val 325	ATT Ile	GTG Val	CAT His	GAA Glu	AAT Asn 330	1011
CCC Pro	TTC Phe	ATC Ile	AGC Ser	GTC Val 335	GAG Glu	TGG Trp	CTC Leu	AAA Lys	GGA Gly 340	CCC Pro	ATC Ile	CTG Leu	GAG Glu	GCC Ala 345	ACG Thr	1059
GCA Ala	GGA Gly	GAC Asp	GAG Glu 350	CTG Leu	GTG Val	AAG Lys	CTG Leu	CCC Pro 355	GTG Val	AAG Lys	CTG Leu	GCA Ala	GCG Ala 360	TAC Tyr	CCC Pro	1107
CCG Pro	CCC Pro	GAG Glu 365	TTC Phe	CAG Gln	TGG Trp	TAC Tyr	AAG Lys 370	GAT Asp	GGA Gly	AAG Lys	GCA Ala	CTG Leu 375	TCC Ser	GGG Gly	CGC Arg	1155
CAC His	AGT Ser 380	CCA Pro	CAT His	GCC Ala	CTG Leu	GTG Val 385	CTC Leu	AAG Lys	GAG Glu	GTG Val	ACA Thr 390	GAG Glu	GCC Ala	AGC Ser	ACA Thr	1203
GGC Gly 395	ACC Thr	TAC Tyr	ACC Thr	CTC Leu	GCC Ala 400	CTG Leu	TGG Trp	AAC Asn	TCC Ser	GCT Ala 405	GCT Ala	GGC Gly	CTG Leu	AGG Arg	CGC Arg 410	1251
AAC Asn	ATC Ile	AGC Ser	CTG Leu	GAG Glu 415	CTG Leu	GTG Val	GTG Val	AAT Asn	GTG Val 420	CCC Pro	CCC Pro	CAG Gln	ATA Ile	CAT His 425	GAG Glu	1299
AAG Lys	GAG Glu	GCC Ala	TCC Ser 430	TCC Ser	CCC Pro	AGC Ser	ATC Ile	TAC Tyr 435	TCG Ser	CGT Arg	CAC His	AGC Ser	CGC Arg 440	CAG Gln	GCC Ala	1347

FIG. 1B

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CTC Leu	ACC Thr	TGC Cys 445	ACG Thr	GCC Ala	TAC Tyr	GGG Gly	GTG Val 450	CCC Pro	CTG Leu	CCT Pro	CTC Leu	AGC Ser 455	ATC Ile	CAG Gln	TGG Trp	1395
CAC His	TGG Trp 460	CGG Arg	CCC Pro	TGG Trp	ACA Thr	CCC Pro 465	TGC Cys	AAG Lys	ATG Met	TTT Phe	GCC Ala 470	CAG Gln	CGT Arg	AGT Ser	CTC Leu	1443
CGG Arg 475	CGG Arg	CGG Arg	CAG Gln	CAG Gln	CAA Gln 480	GAC Asp	CTC Leu	ATG Met	CCA Pro	CAG Gln 485	TGC Cys	CGT Arg	GAC Asp	TGG Trp	AGG Arg 490	1491
GCG Ala	GTG Val	ACC Thr	ACG Thr	CAG Gln 495	GAT Asp	GCC Ala	GTG Val	AAC Asn	CCC Pro 500	ATC Ile	GAG Glu	AGC Ser	CTG Leu	GAC Asp 505	ACC Thr	1539
TGG Trp	ACC Thr	GAG Glu	TTT Phe 510	GTG Val	GAG Glu	GGA Gly	AAG Lys	AAT Asn 515	AAG Lys	ACT Thr	GTG Val	AGC Ser	AAG Lys 520	CTG Leu	GTG Val	1587
ATC Ile	CAG Gln	AAT Asn 525	GCC Ala	AAC Asn	GTG Val	TCT Ser	GCC Ala 530	ATG Met	TAC Tyr	AAG Lys	TGT Cys	GTG Val 535	GTC Val	TCC Ser	AAC Asn	1635
AAG Lys	GTG Lys 540	GGC Gly	CAG Gln	GAT Asp	GAG Glu	CGG Arg 545	CTC Leu	ATC Ile	TAC Tyr	TTC Phe	TAT Tyr 550	GTG Val	ACC Thr	ACC Thr	ATC Ile	1683
CCC Pro 555	GAC Asp	GGC Gly	TTC Phe	ACC Thr	ATC Ile 560	GAA Glu	TCC Ser	AAG Lys	CCA Pro	TCC Ser 565	GAG Glu	GAG Glu	CTA Leu	CTA Leu	GAG Glu 570	1731
GGC Gly	CAG Gln	CCG Pro	GTG Val	CTC Leu 575	CTG Leu	AGC Ser	TGC Cys	CAA Gln 580	GCC Ala 580	GAC Asp	AGC Ser	TAC Tyr	AAG Lys	TAC Tyr 585	GAG Glu	1779
CAT His	CTG Leu	CGC Arg	TGG Trp 590	TAC Tyr	CGC Arg	CTC Leu	AAC Asn	CTG Leu 595	TCC Ser	ACG Thr	CTG Leu	CAC His	GAT Asp 600	GCG Ala	CAC His	1827
GGG Gly	AAC Asn	CCG Pro 605	CTT Leu	CTG Leu	CTC Leu	GAC Asp	TGC Cys 610	AAG Lys	AAC Asn	GTG Val	CAT His	CTG Leu 615	TTC Phe	GCC Ala	ACC Thr	1875
CCT Pro	CTG Leu 620	GCC Ala	GCC Ala	AGC Ser	CTG Leu	GAG Glu 625	GAG Glu	GTG Val	GCA Ala	CCT Pro	GGG Gly 630	GCG Ala	CGC Arg	CAC His	GCC Ala	1923
ACG Thr 635	CTC Leu	AGC Ser	CTG Leu	AGT Ser	ATC Ile 640	CCC Pro	CGC Arg	GTC Val	GCG Ala	CCC Pro 645	GAG Glu	CAC His	GAG Glu	GGC Gly	CAC His 650	1971
TAT Tyr	GTG Val	TGC Cys	GAA Glu	GTG Val 655	CAA Gln	GAC Asp	CGG Arg	CGC Arg	AGC Ser 660	CAT His	GAC Asp	AAG Lys	CAC His	TGC Cys 665	CAC His	2019

FIG. 1C

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AAG Lys	AAG Lys	TAC Tyr	CTG Leu 670	TCG Ser	GTG Val	CAG Gln	GCC Ala	CTG Leu 675	GAA Glu	GCC Ala	CCT Pro	CGG Arg	CTC Leu 680	ACG Thr	CAG Gln	2067
AAC Asn	TTG Leu	ACC Thr 685	GAC Asp	CTC Leu	CTG Leu	GTG Val	AAC Asn 690	GTG Val	AGC Ser	GAC Asp	TCG Ser	CTG Leu 695	GAG Glu	ATG Met	CAG Gln	2115
TGC Cys	TTG Leu 700	GTG Val	GCC Ala	GGA Gly	GCG Ala	CAC His 705	GCG Ala	CCC Pro	AGC Ser	ATC Ile	GTG Val 710	TGG Trp	TAC Tyr	AAA Lys	GAC Asp	2163
GAG Glu 715	AGG Arg	CTG Leu	CTG Leu	GAG Glu	GAA Glu 720	AAG Lys	TCT Ser	GGA Gly	GTC Val	GAC Asp 725	TTG Leu	GCG Ala	GAC Asp	TCC Ser	AAC Asn 730	2211
CAG Gln	AAG Lys	CTG Leu	AGC Ser	ATC Ile 735	CAG Gln	CGC Arg	GTG Val	GCG Arg	GAG Glu 740	GAG Glu	GAT Asp	GCG Ala	GGA Gly	CCG Pro 745	TAT Tyr	2259
CTG Leu	TGC Cys	AGC Ser	GTG Val 750	TGC Cys	AGA Arg	CCC Pro	AAG Lys	GGC Gly 755	TGC Cys	GTC Val	AAC Asn	TCC Ser	TCC Ser 760	GCC Ala	AGC Ser	2307
GTG Val	GCC Ala	GTG Val 765	GAA Glu	GGC Gly	TCC Ser	GAG Glu	GAT Asp 770	AAG Lys	GGC Gly	AGC Ser	ATG Met	GAG Glu 775	ATC Ile	GTG Val	ATC Ile	2355
CTT Leu	GTC Val 780	GGT Gly	ACC Thr	GGC Gly	GTC Val	ATC Ile 785	GCT Ala	GTC Val	TTC Phe	TTC Phe	TGG Trp 790	GTC Val	CTC Leu	CTC Leu	CTC Leu	2403
CTC Leu 795	ATC Ile	TTC Phe	TGT Cys	AAC Asn	ATG Met 800	AGG Arg	AGG Arg	CCG Pro	GCC Ala	CAC His 805	GCA Ala	GAC Asp	ATC Ile	AAG Lys	ACG Thr 810	2451
GGC Gly	TAC Tyr	CTG Leu	TCC Ser	ATC Ile 815	ATC Ile	ATG Met	GAC Asp	CCC Pro	GGG Gly 820	GAG Glu	GTG Val	CCT Pro	CTG Leu	GAG Glu 825	GAG Glu	2499
CAA Gln	TGC Cys	GAA Glu	TAC Tyr 830	CTG Leu	TCC Ser	TAC Tyr	GAT Asp	GCC Ala 835	AGC Ser	CAG Gln	TGG Trp	GAA Glu	TTC Phe 840	CCC Pro	CGA Arg	2547
GAG Glu	CGG Arg	CTG Leu 845	CAC His	CTG Leu	GGG Gly	AGA Arg	GTG Val 850	CTC Leu	GGC Gly	TAC Tyr	GGC Gly	GCC Ala 855	TTC Phe	GGG Gly	AAG Lys	2595
GTG Val	GTG Val 860	GAA Glu	GCC Ala	TCC Ser	GCT Ala	TTC Phe 865	GGC Gly	ATC Ile	CAC His	AAG Lys	GGC Gly 870	AGC Ser	AGC Ser	TGT Cys	GAC Asp	2643
ACC Thr 875	GTG Val	GCC Ala	GTG Val	AAA Lys	ATG Met 880	CTG Leu	AAA Lys	GAG Glu	GGC Gly	GCC Ala	ACG Thr	GCC Ala	AGC Ser	GAG Glu	CAG Gln 890	2691

FIG. 1D

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CGC	GCG	CTG	ATG	TCG	GAG	CTC	AAG	ATC	CTC	ATT	CAC	ATC	GGC	AAC	CAC	2739
Arg	Ala	Leu	Met	Ser	Glu	Leu	Lys	Ile	Leu	Ile	His	Ile	Gly	Asn	His	895
CTC	AAC	GTG	GTC	AAC	CTC	CTC	GGG	GCG	TGC	ACC	AAG	CCG	CAG	GGC	CCC	2787
Leu	Asn	Val	Val	Asn	Leu	Leu	Gly	Ala	Cys	Thr	Lys	Pro	Gln	Gly	Pro	910
CTC	ATG	GTG	ATC	GTG	GAG	TTC	TGC	AAG	TAC	GGC	AAC	CTC	TCC	AAC	TTC	2835
Leu	Met	Val	Ile	Val	Glu	Phe	Cys	Lys	Tyr	Gly	Asn	Leu	Ser	Asn	Phe	925
CTG	CGC	GCC	AAG	CGG	GAC	GCC	TTC	AGC	CCC	TGC	GCG	GAG	AAG	TCT	CCC	2883
Leu	Arg	Ala	Lys	Arg	Asp	Ala	Phe	Ser	Pro	Cys	Ala	Glu	Lys	Ser	Pro	940
GAG	CAG	CGC	GGA	CGC	TTC	CGC	GCC	ATG	GTG	GAG	CTC	GCC	AGG	CTG	GAT	2931
Glu	Gln	Arg	Gly	Arg	Phe	Arg	Ala	Met	Val	Glu	Leu	Ala	Arg	Leu	Asp	955
CGG	AGG	CGG	CCG	GGG	AGC	AGC	GAC	AGG	GTC	CTC	TTC	GCG	CGG	TTC	TCG	2979
Arg	Arg	Arg	Pro	Gly	Ser	Ser	Asp	Arg	Val	Leu	Phe	Ala	Arg	Phe	Ser	975
AAG	ACC	GAG	GGC	GGA	GCG	AGG	CGG	GCT	TCT	CCA	GAC	CAA	GAA	GCT	GAG	3027
Lys	Thr	Glu	Gly	Gly	Ala	Arg	Arg	Ala	Ser	Pro	Asp	Gln	Glu	Ala	Glu	990
GAC	CTG	TGG	CTG	AGC	CCG	CTG	ACC	ATG	GAA	GAT	CTT	GTC	TGC	TAC	AGC	3075
Asp	Leu	Trp	Leu	Ser	Pro	Leu	Thr	Met	Glu	Asp	Leu	Val	Cys	Tyr	Ser	1005
TTC	CAG	GTG	GCC	AGA	GGG	ATG	GAG	TTC	CTG	GCT	TCC	CGA	AAG	TGC	ATC	3123
Phe	Gln	Val	Ala	Arg	Gly	Met	Glu	Phe	Leu	Ala	Ser	Arg	Lys	Cys	Ile	1020
CAC	AGA	GAC	CTG	GCT	GCT	CGG	AAC	ATT	CTG	CTG	TCG	GAA	AGC	GAC	GTG	3171
His	Arg	Asp	Leu	Ala	Ala	Arg	Asn	Ile	Leu	Leu	Ser	Glu	Ser	Asp	Val	1035
GTG	AAG	ATC	TGT	GAC	TTT	GGC	CTT	GCC	CGG	GAC	ATC	TAC	AAA	GAC	CCC	3219
Val	Lys	Ile	Cys	Asp	Phe	Gly	Leu	Ala	Arg	Asp	Ile	Tyr	Lys	Asp	Pro	1040
GAC	TAC	GTC	CGC	AAG	GGC	AGT	GCC	CGG	CTG	CCC	CTG	AAG	TGG	ATG	GCC	3267
Asp	Tyr	Val	Arg	Lys	Gly	Ser	Ala	Arg	Leu	Pro	Leu	Lys	Trp	Met	Ala	1055
CCT	GAA	AGC	ATC	TTC	GAC	AAG	GTG	TAC	ACC	ACG	CAG	AGT	GAC	GTG	TGG	3315
Pro	Glu	Ser	Ile	Phe	Asp	Lys	Val	Tyr	Thr	Thr	Gln	Ser	Asp	Val	Trp	1070
TCC	TTT	GGG	GTG	CTT	CTC	TGG	GAG	ATC	TTC	TCT	CTG	GGG	GCC	TCC	CCG	3363
Ser	Phe	Gly	Val	Leu	Leu	Trp	Glu	Ile	Phe	Ser	Leu	Gly	Ala	Ser	Pro	1100

FIG. 1E

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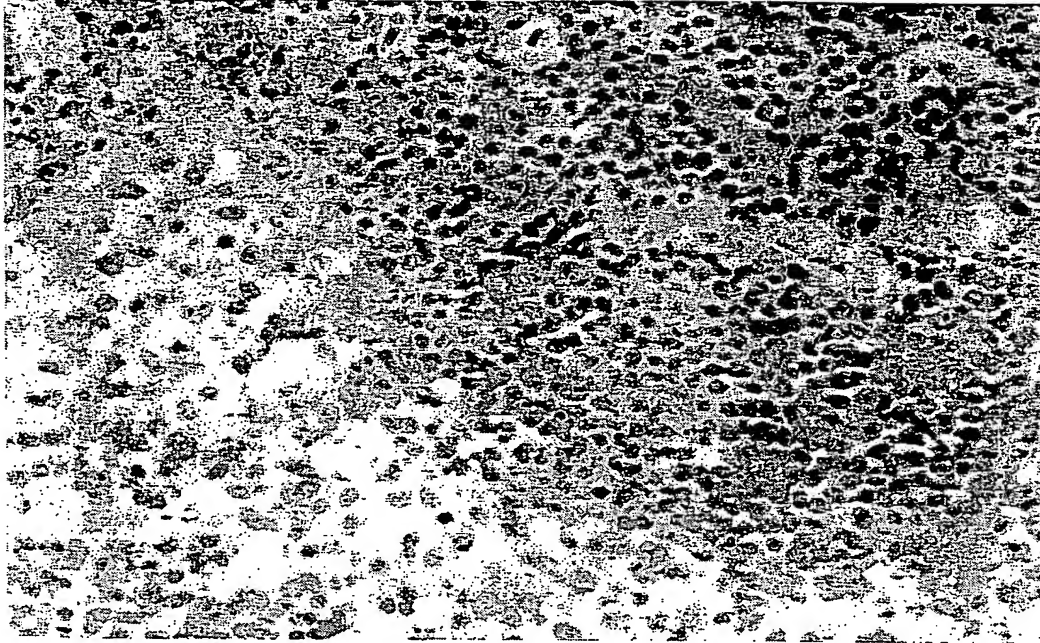
TAC Tyr 1115	CCT Pro	GGG Gly	GTG Val	CAG Gln 1120	ATC Ile	AAT Asn	GAG Glu	GAG Glu	TTC Phe 1125	TGC Cys	CAG Gln	CGC Arg	GTG Val	AGA Arg 1130	GAC Asp	3411
GGC Gly	ACA Thr	AGG Arg	ATG Met 1135	AGG Arg	GCC Ala	CCG Pro	GAG Glu	CTG Leu 1140	GCC Ala	ACT Thr	CCC Pro	GCC Ala	ATA Ile 1145	CGC Arg	CAC His	3459
ATC Ile	ATG Met	CTG Leu 1150	AAC Asn	TGC Cys	TGG Trp	TCC Ser	GGA Gly 1155	GAC Asp	CCC Pro	AAG Lys	GCG Ala	AGA Arg 1160	CCT Pro	GCA Ala	TTC Phe	3507
TGC Ser	GAC Asp 1165	CTG Leu	GTG Val	GAG Glu	ATC Ile	CTG Leu 1170	GGG Gly 1175	GAC Asp	CTG Leu	CTC Leu	CAG Gln 1175	GGC Gly	AGG Arg	GGC Gly	CTG Leu	3555
CAA Gln 1180	GAG Glu	GAA Glu	GAG Glu	GAG Glu	GTC Val 1185	TGC Cys	ATG Met	GCC Ala	CCG Pro	CGC Arg 1190	AGC Ser	TCT Ser	CAG Gln	AGA Ser	TCA Ser	3603
GAA Glu 1195	GAG Glu	GGC Gly	AGC Ser	TTC Phe 1200	TCG Ser	CAG Gln	GTG Val	TCC Ser	ACC Thr 1205	ATG Met	GCC Ala	CTA Leu	CAC His	ATC Ile 1210	GCC Ala	3651
CAG Gln	GCT Ala	GAC Asp 1215	GCT Ala	GAG Glu	GAC Asp	AGC Ser	CCG Pro	CCA Pro 1220	AGC Ser	CTG Leu	CAG Gln	CGC Arg	CAC His 1225	AGC Ser	CTG Leu	3699
GCC Ala	GCC Ala	AGG Arg 1230	TAT Tyr	TAC Tyr	AAC Asn	TGG Trp	GTG Val 1235	TCC Ser	TTT Phe	CCC Pro	GGG Gly	TGC Cys 1240	CTG Leu	GCC Ala	AGA Arg	3747
GGG Gly	GCT Ala 1245	GAG Glu	ACC Thr	CGT Arg	GGT Gly	TCC Ser 1250	TCC Ser	AGG Arg	ATG Met	AAG Lys	ACA Thr 1255	TTT Phe	GAG Glu	GAA Glu	TTC Phe	3795
CCC Phe 1260	ATG Met	ACC Thr	CCA Pro	ACG Thr	ACC Thr 1265	TAC Tyr	AAA Lys	GGC Gly	TCT Ser 1270	GTG Val	GAC Asp 1270	AAC Asn	CAG Gln	ACA Thr	GAC Asp	3843
AGT Ser 1275	GGG Gly	ATG Met	GTG Val	CTG Leu 1280	GCC Ala	TCG Ser	GAG Glu	GAG Glu	TTT Phe 1285	GAG Glu	CAG Gln	ATA Ile	GAG Glu	AGC Ser 1290	AGG Arg	3891
CAT His	AGA Arg	CAA Gln 1295	GAA Glu	AGC Ser	GGC Gly	TTC Phe	AGG Arg	TAGCTGAAGC AGAGAGAGAG AAGGCAGCAT								3945
ACGTCAGCAT TTTCTTCTCT GCACTTATAA GAAAGATCAA AGACTTTAAG ACTTTTCGCTA 4005																
TTTCTTCTAC TGCTATCTAC TACAACTTC AAAGAGGAAC CAGGAGGACA AGAGGAGCAT 4065																
GAAAGTGGAC AAGGAGTGTG ACCACTGAAG CACCACAGGG AGGGGTTAGG CCTCCGGATG 4125																
ACTGCGGGCA GGCCTGGATA ATATCCAGCC TCCCACAAGA AGCTGGTGGA GCAGAGTGTT 4185																
CCCTGACTCC TCCAAGGAAA GGGAGACGCC CTTTCATGGT CTGCTGAGTA ACAGGTGCCT 4245																
TCCCAGACAC TGGCGTTACT GCTTGACCAA AGAGCCCTCA AGCGGCCCTT ATGCCAGCGT 4305																
GACAGAGGGC TCACCTCTTG CTTTCTAGGT CACTTCTCAC AATGTCCCTT CAGCACCTGA 4365																
CCCTGTGCCC GCCGATTATT CTTTGTAAT ATGAGTAATA CATCAAAGAG TAGTATTAAA 4425																
AGCTAATTAA TCATGTTTAT AAAAAA 4450																

FIG. 1F

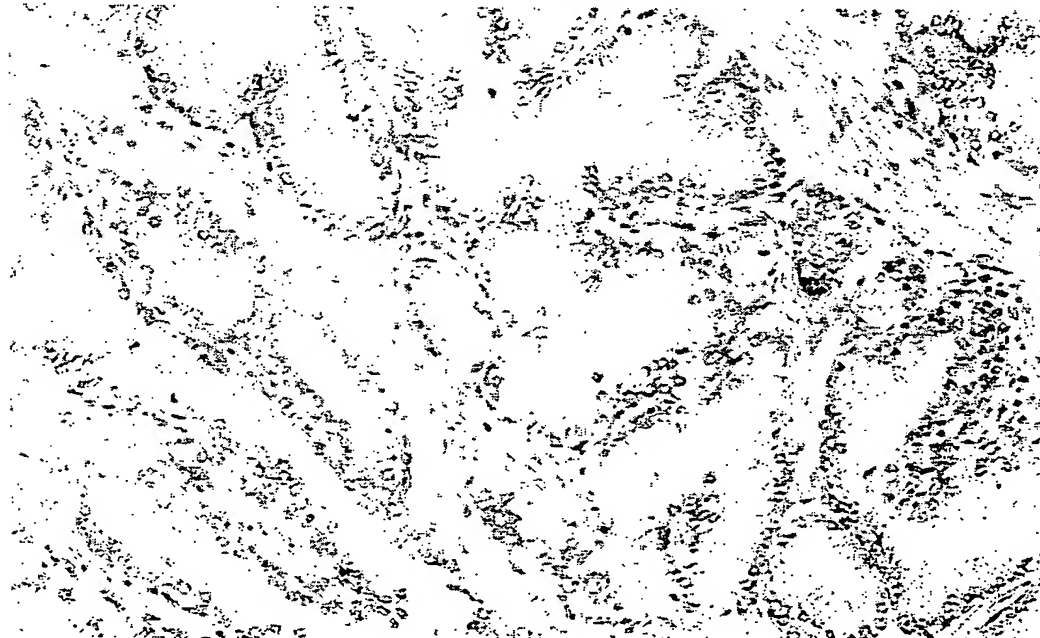
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Benign Node stained with anti-FLT-4 antibody:

**FIG.2A**

BPH stained with anti-FLT-4:

**FIG.2B**

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Node with prostatic metastases stained with anti-FLT-4 antibody:

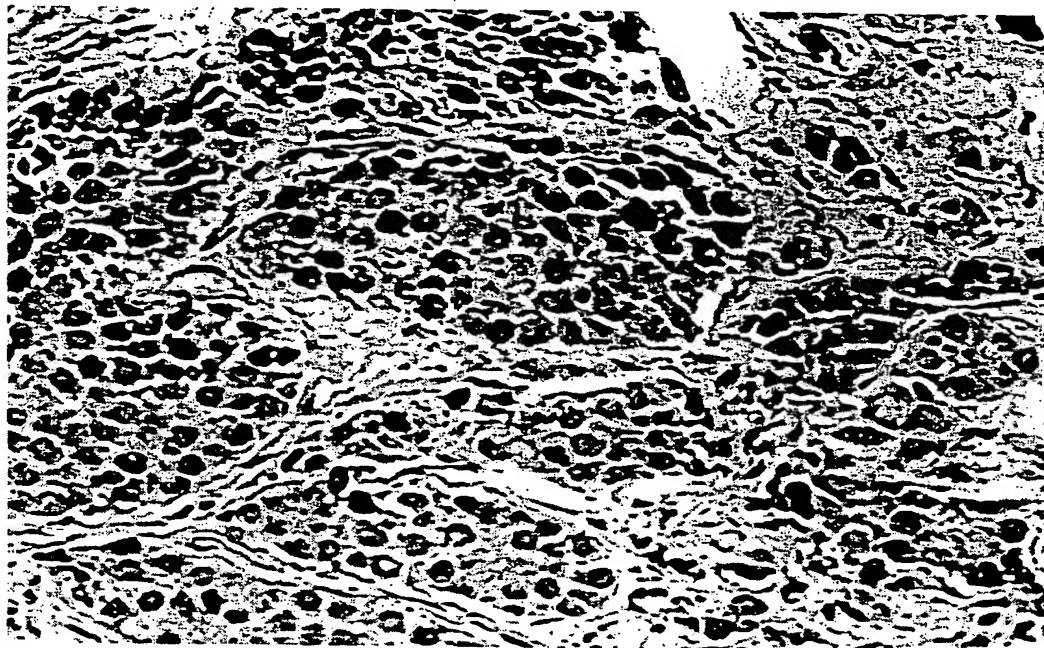


FIG.2C

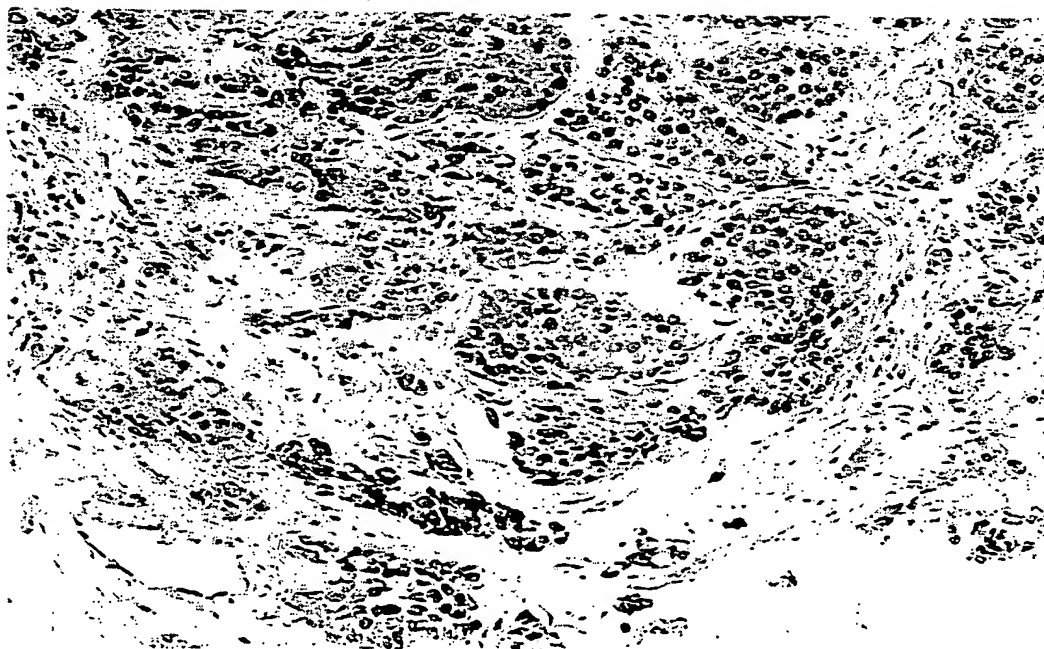


FIG.2D

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BPH and extensive CoP stained with anti-FLT-4 showing positive reactivity in CoP and mostly negative reactivity in BPH:

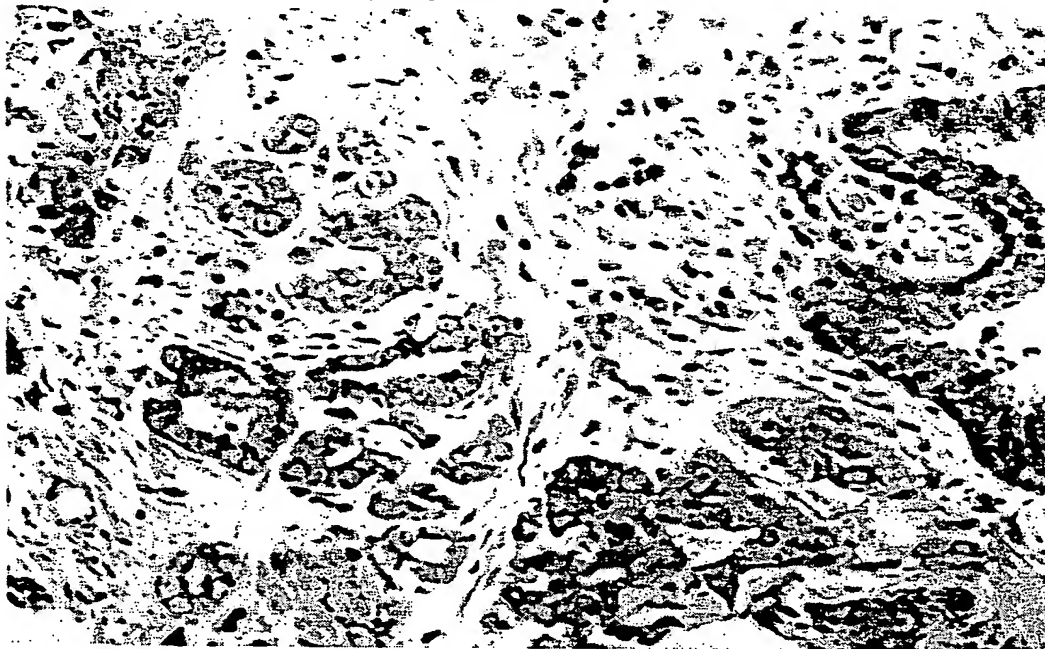


FIG.2E

Extensive CoP stained with anti-FLT-4:

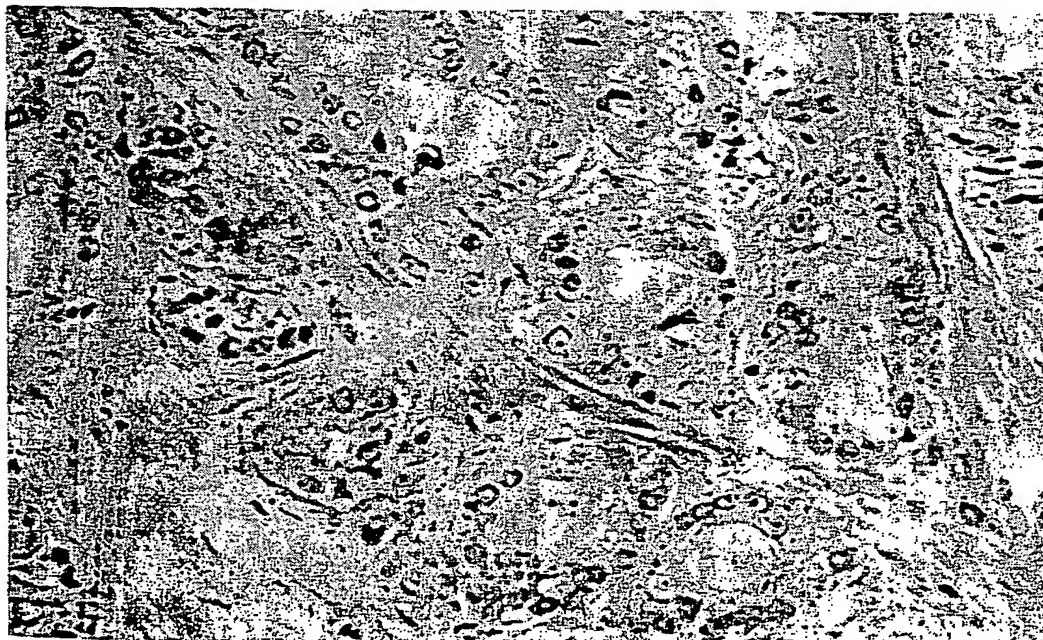


FIG.2F

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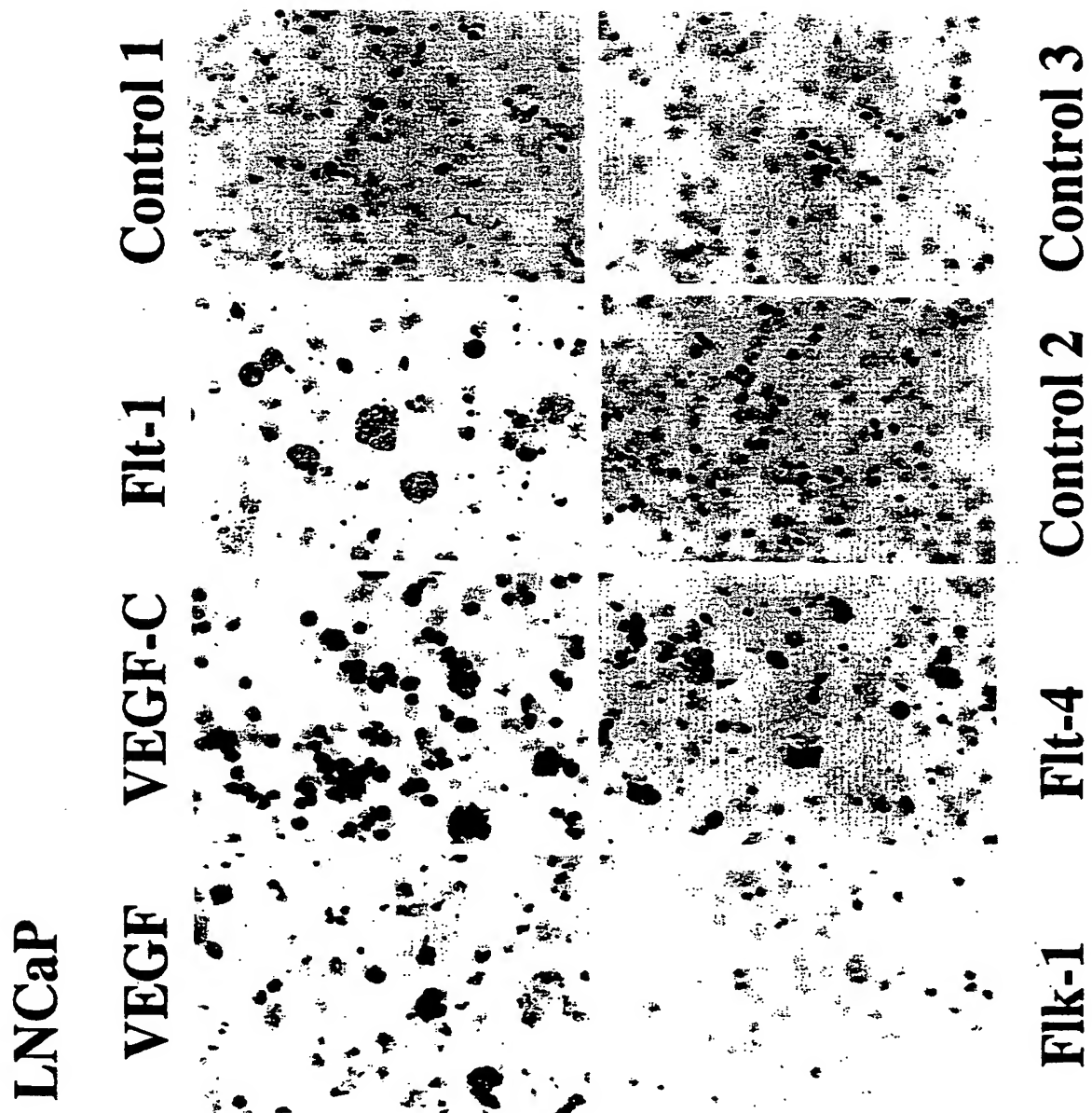


FIG. 3A

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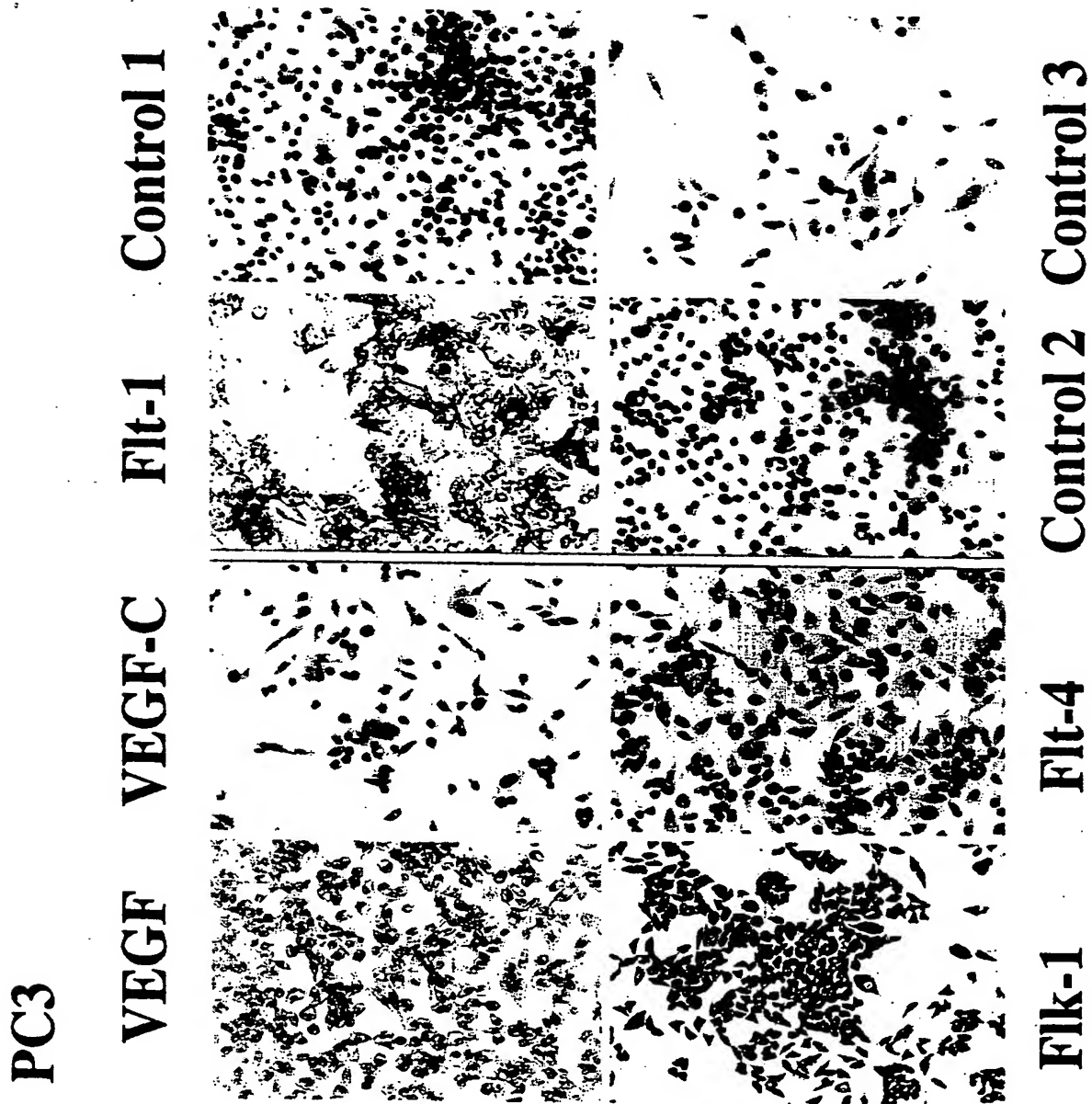


FIG.3B

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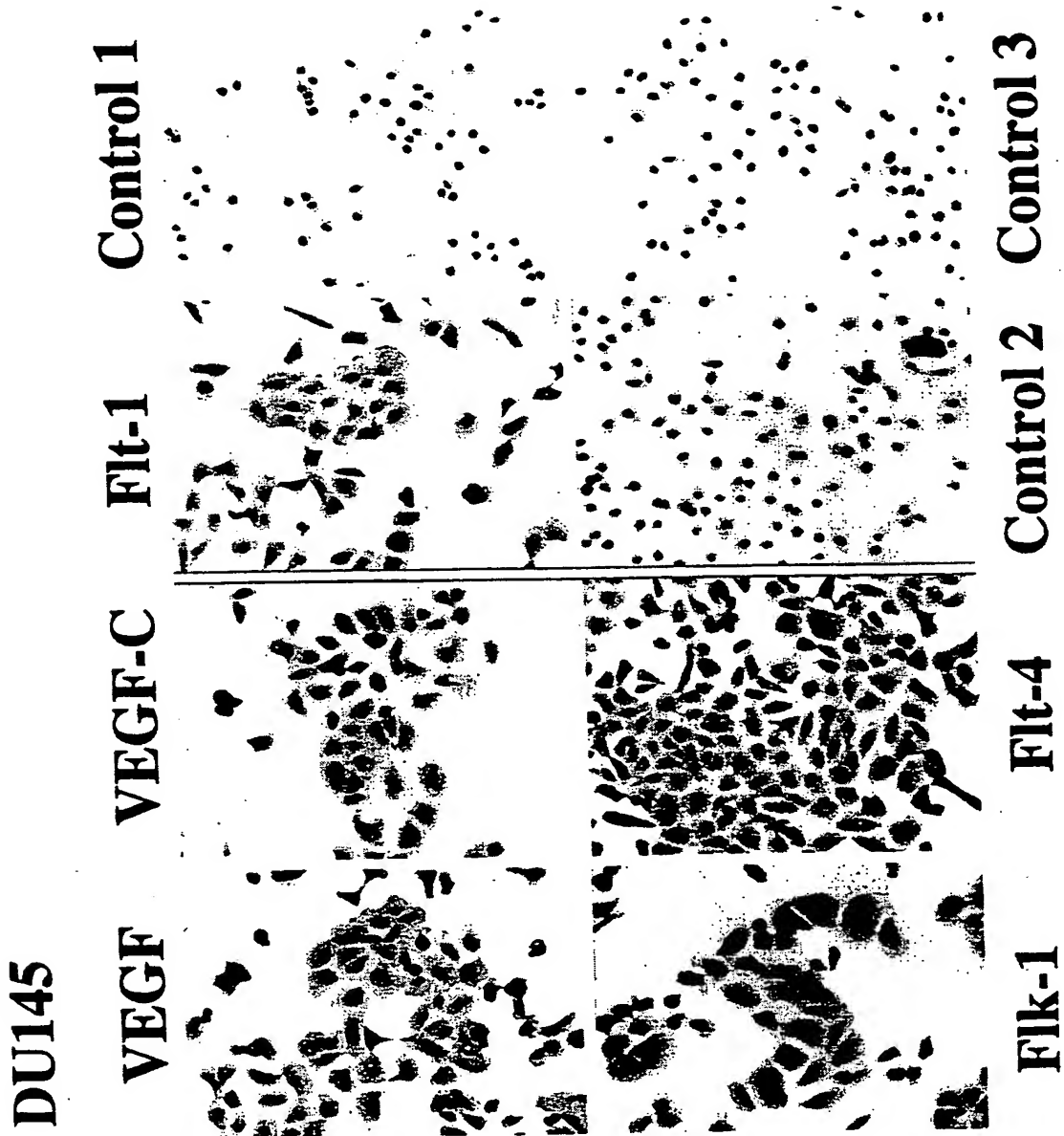


FIG.3C

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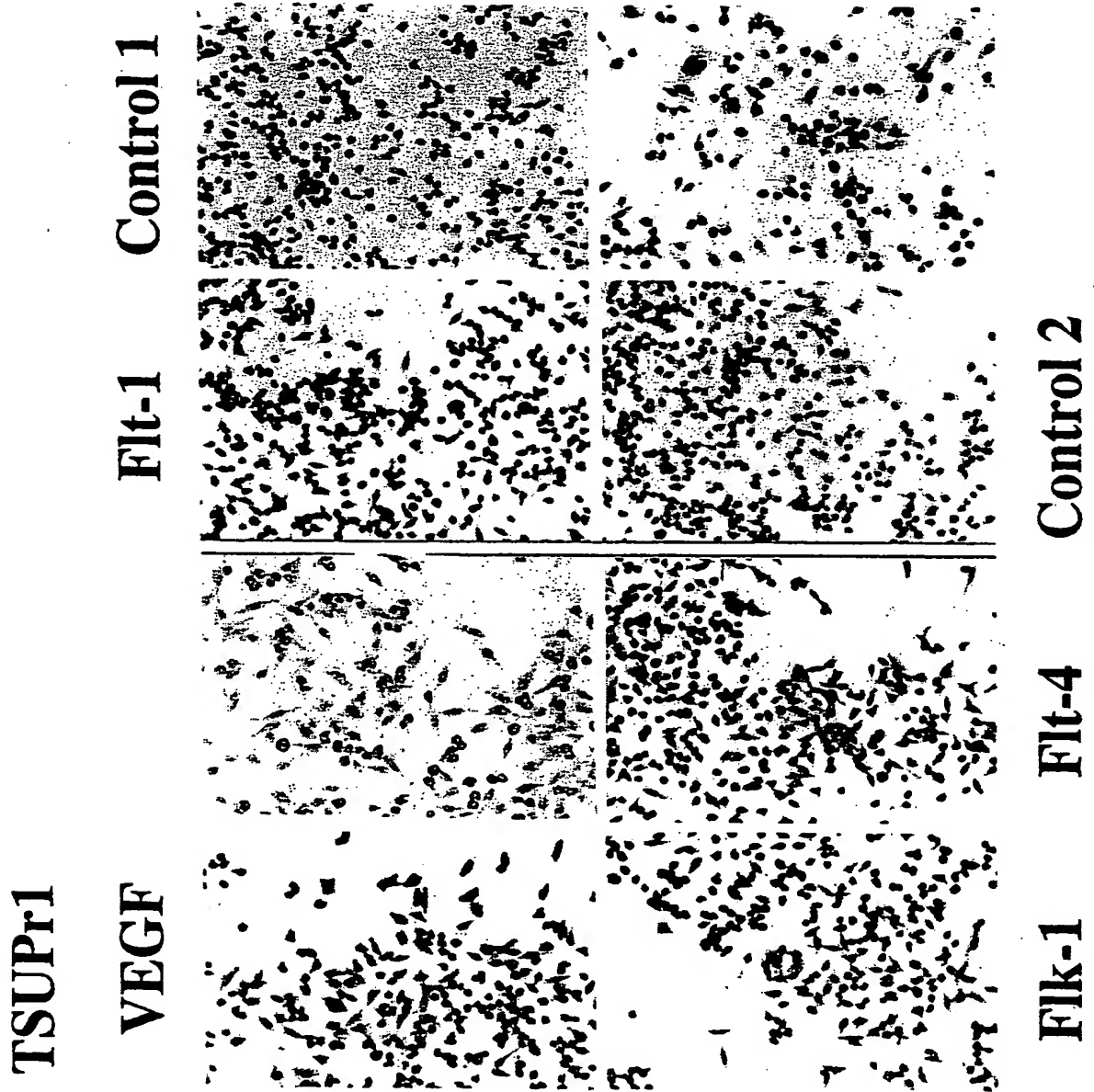


FIG.3D

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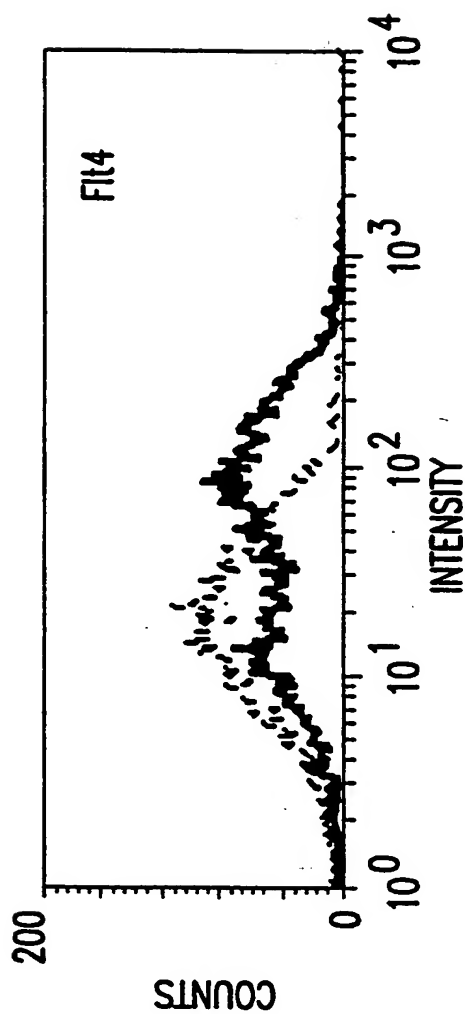
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VEGF-C	VEGF	FLK-1	FLT-4	β -Actin
LNCaP	LNCaP	LNCaP	LNCaP	LNCaP
PC3	PC3	PC3	PC3	PC3
DU145	DU145	DU145	DU145	DU145
TSUP-r1	TSUP-r1	TSUP-r1	TSUP-r1	TSUP-r1
H2O	H2O	H2O	H2O	H2O

FIG. 4

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The dotted line shows the isotype control while the solid line shows binding with specific antibodies.

FIG.5

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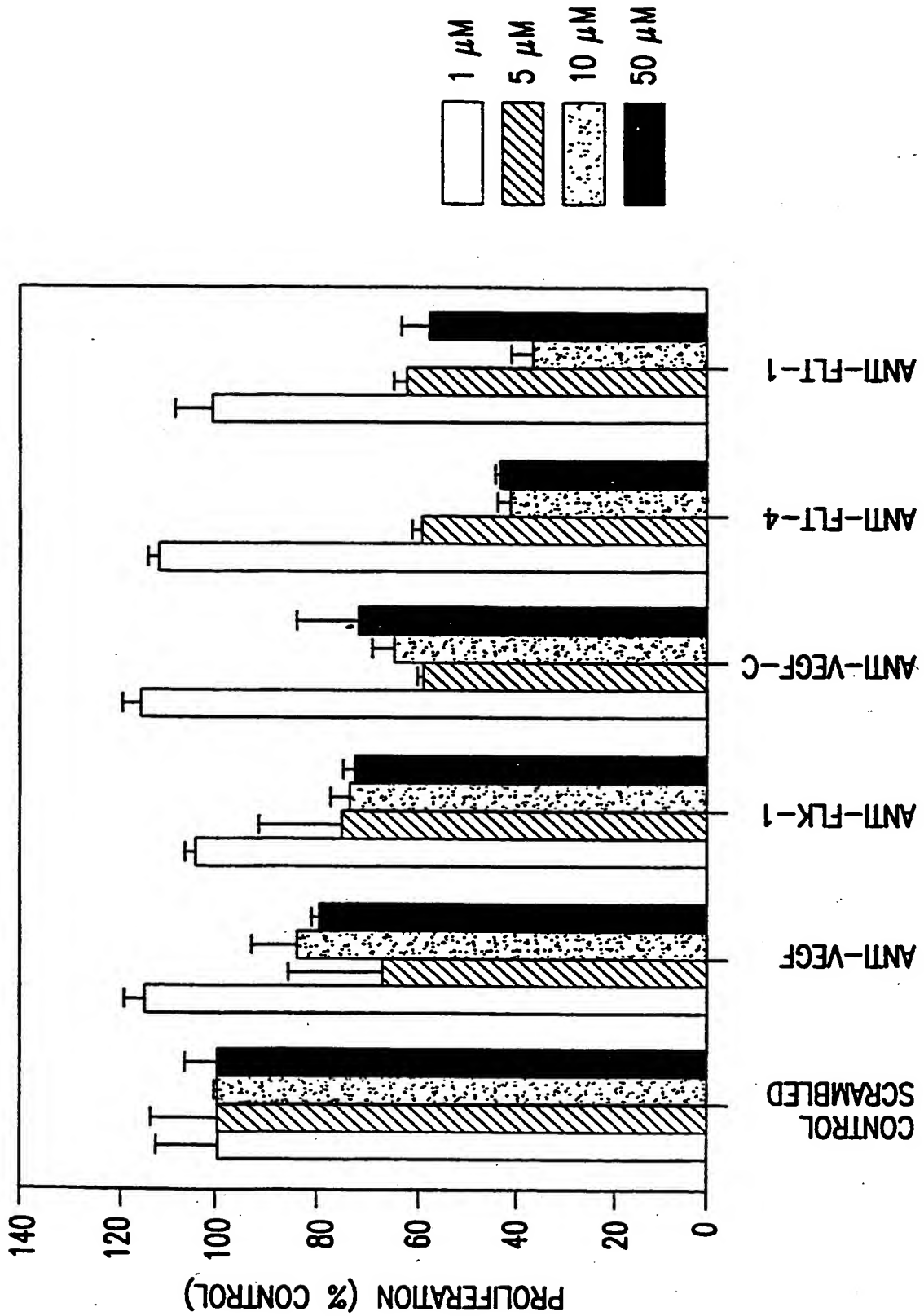


FIG.6

ANTISENSE OLIGONUCLEOTIDE

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